

IMMUNOCHROMATOGRAPHIC METHOD:

# LATERAL FLOW STRIP TEST

Wanwisa Poonlapdecha, Ph.D  
Qualified Diagnostic Development Center (QDD),  
Chulalongkorn University

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เหมาะสมกับชนิดของตัวอย่าง และสารที่ต้องการตรวจสอบ
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# When do you need to develop a new rapid test?

## Design and Development Business Plan

- To generate idea for potential medical devices should be.

### 1 Market Research

- To provide summarised overviews of existing technologies, competitor devices, and economic considerations.

#### Target:

- Identify your problem and set a goal/outcome for the potential intervention.
- Suggest solution(s) to reach the suggested goal/outcome.

#### Methodologies:

- Questionnaires
- Observations
- Face-to-face interviews
- Overview of data collection tools
- Cost-benefit tool
- SWOT-analysis tool
- Strategy tool PEST (Politic, Economic, Socio-Cultural and Technological-factors)
- Porter's Five Forces - Competitor's analysis
- Market Mix (4Ps)

### 2 Project Timeline and Costs

- To define timeframes, budget and solution for each stage

### 3 Prototyping

- Prototype will be the basis for feasibility studies to determine whether you should proceed with your device development project.

### 4 Regulatory Planning

- Determine device classification, which in turn can determine the level of documentation required in each market.
- Classification is based on the risk profile of the device

3 options required for analysis of:-

1. Market feasibility
2. Financial feasibility
3. Regulatory feasibility



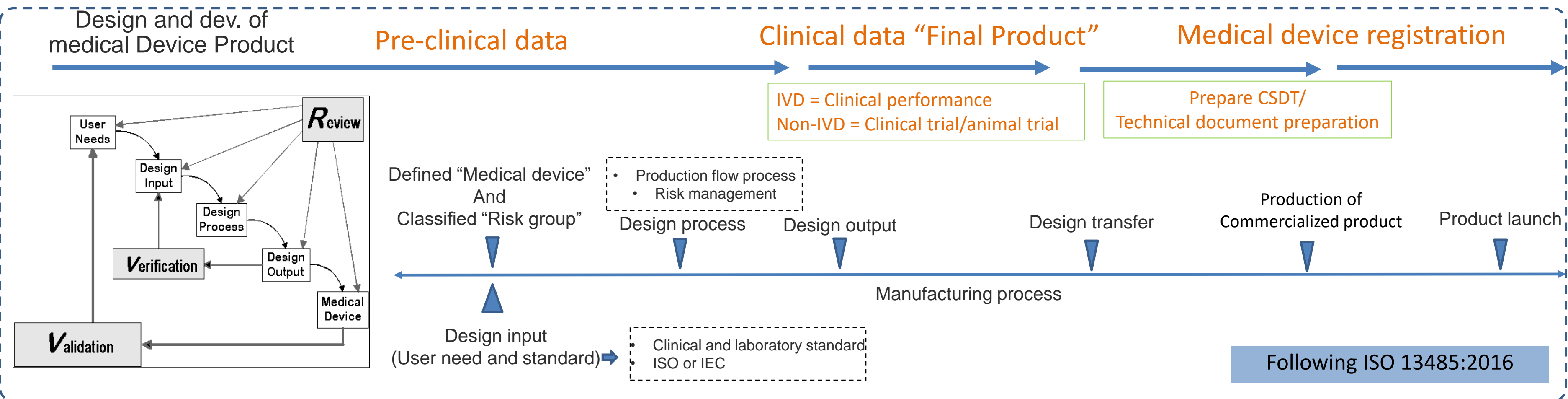
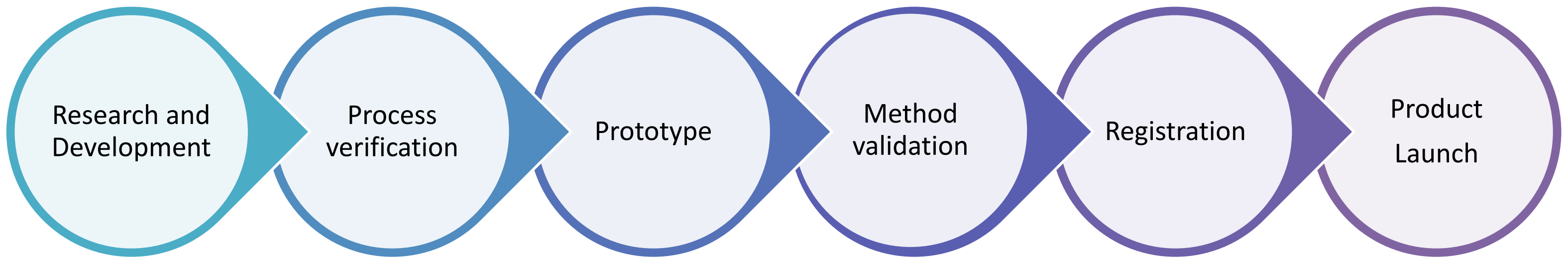
www.bsmarketsci.com

### ชุดทดสอบสเตียรอยด์ (หลักการ TLC)

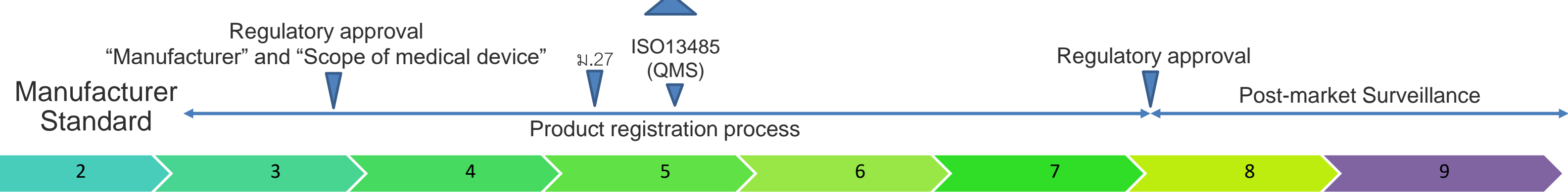


# How to Drive Research product to the Market

6 Steps to Drive



Regulation vs. Research Level



# IMMUNOCHROMATOGRAPHIC METHOD: LATERAL FLOW STRIP TEST

- A paper-based platform to detect analyte in sample
- Based on antigen-antibody interaction
- Capillary force moving
- Applied to detect both qualitative and semi-quantitative
- Getting results in 10-20 mins



- Can be applied for:-
  - : Home testing
  - : Point of care /or
  - : Filed testing

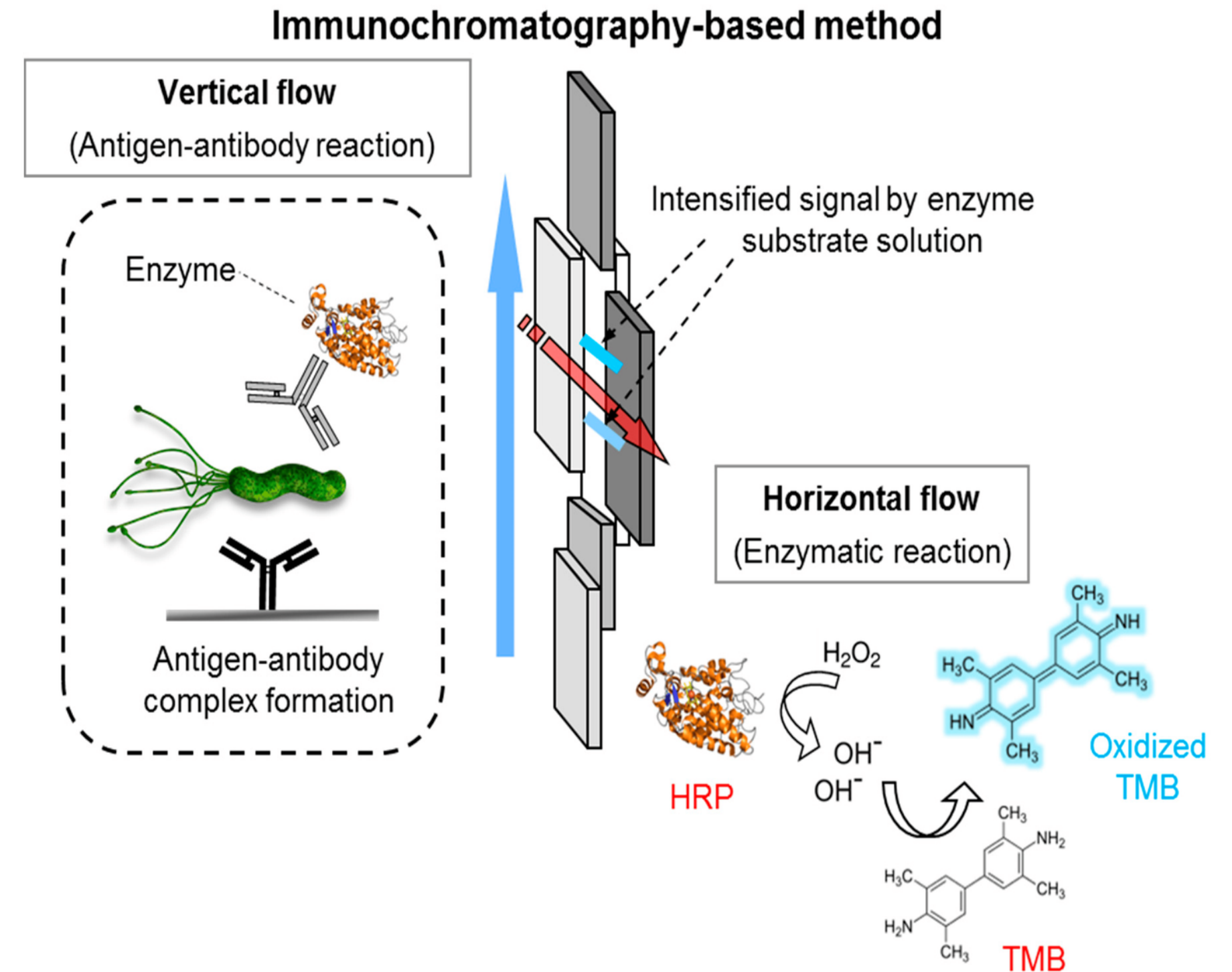


Figure 1 Chromatographic principle: schematic illustration of colorimetric analytical method associated with enzyme-based immunochromatography (e.g., horseradish peroxidase) for onsite testing (Cho and Ku, 2017).

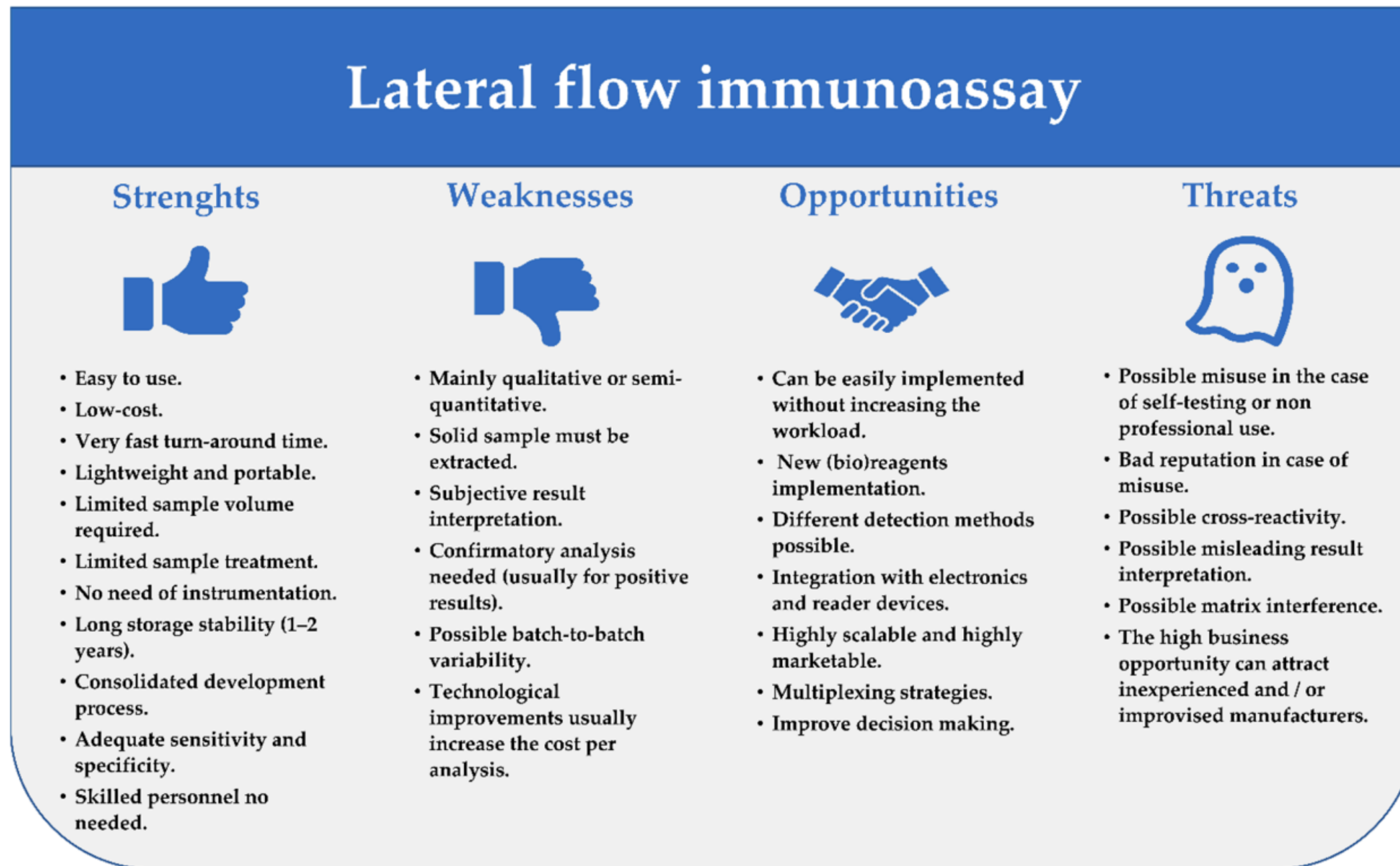
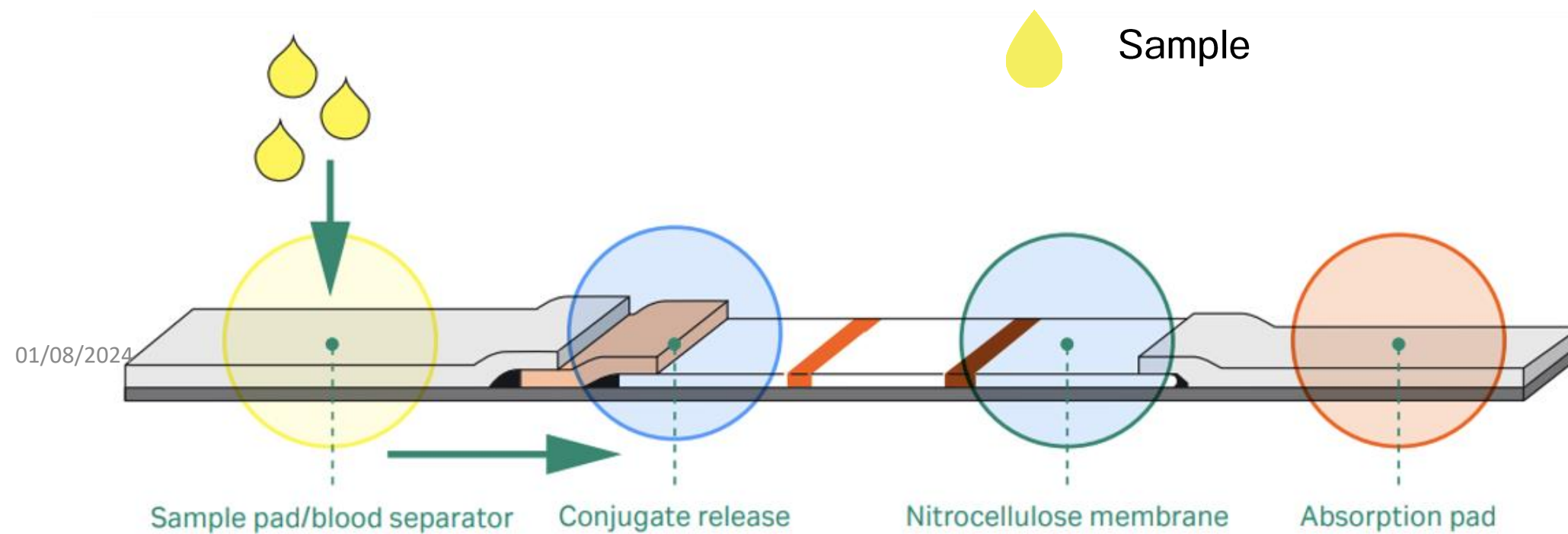


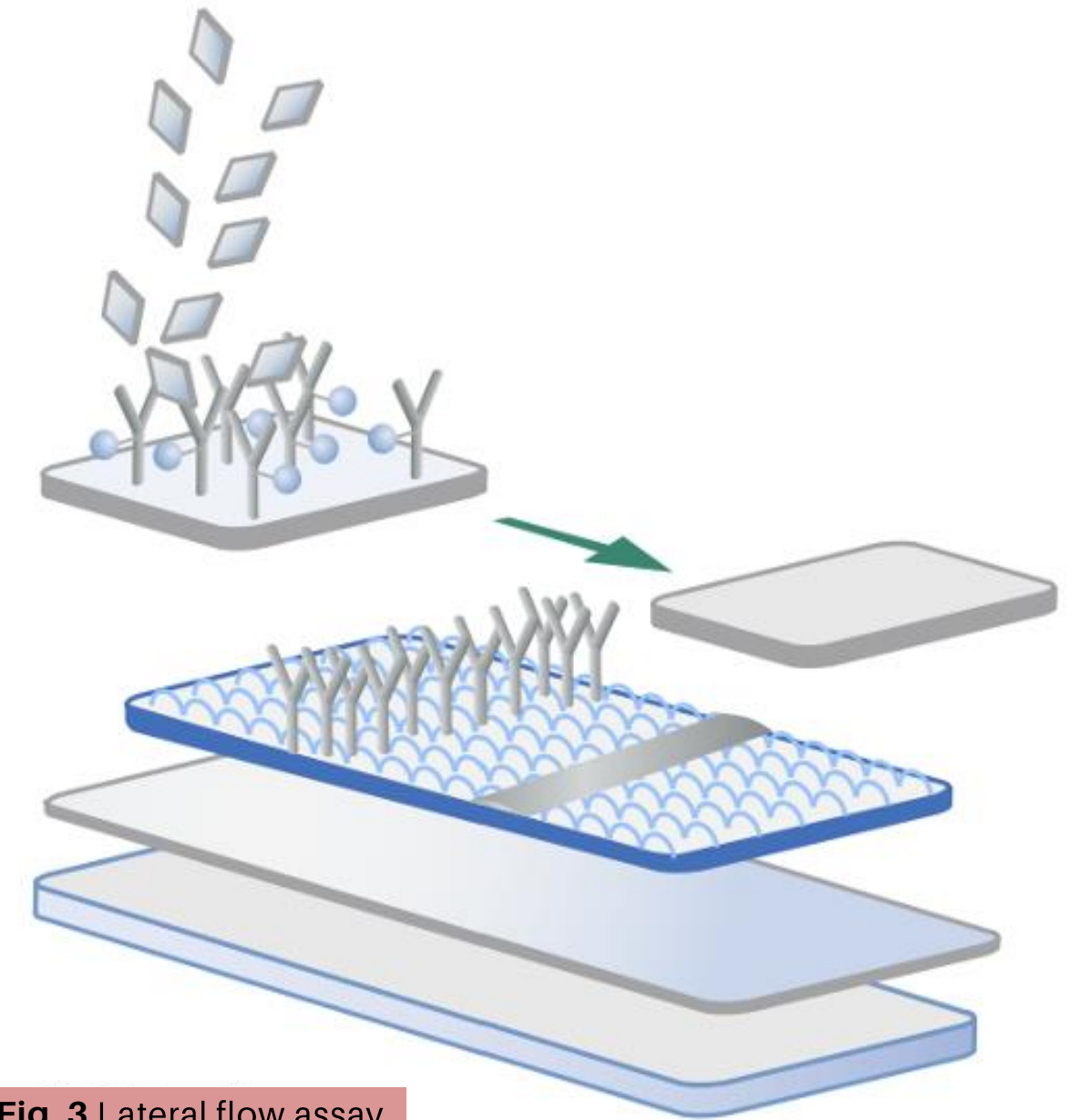
Figure 1. SWOT analysis of the LFIA technique considering its inherent features.

# IMMUNOCHROMATOGRAPHIC METHOD: LATERAL FLOW IMMUNOASSAY

The sample flows through the device and comes in contact with immobilized reagents. The antibody and analyte migrate to a capture zone of membrane-immobilized antibody. Any unreacted tagged antibody flows past the capture zone to absorbent.



**Fig. 2** Drawing of a strip test immunoassay

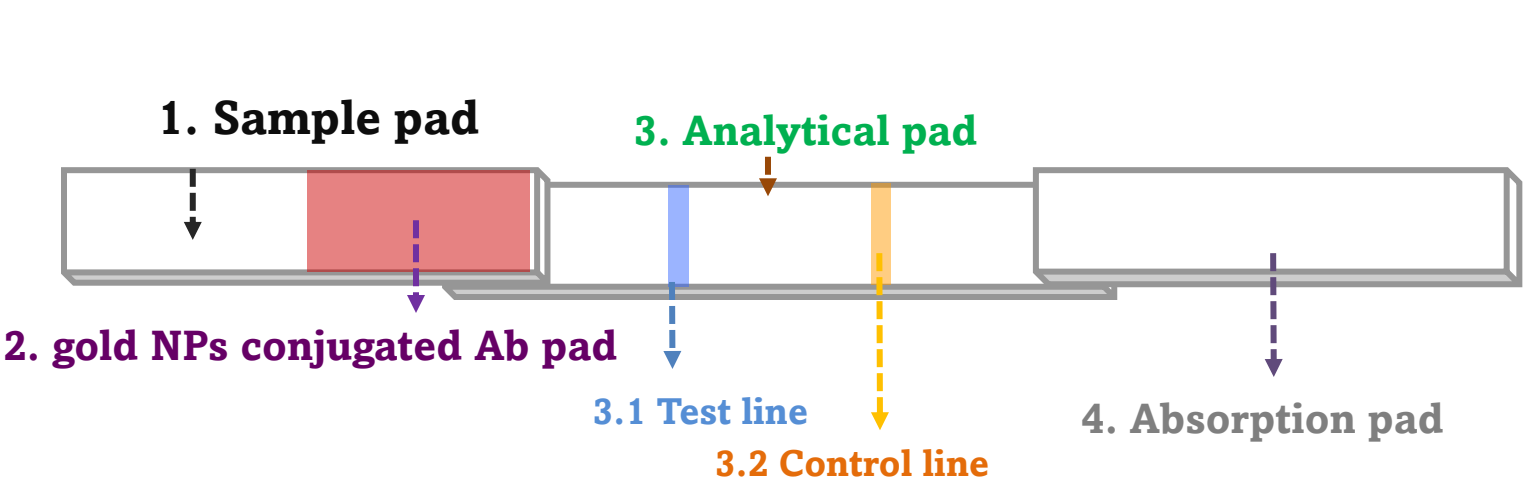


**Fig. 3** Lateral flow assay

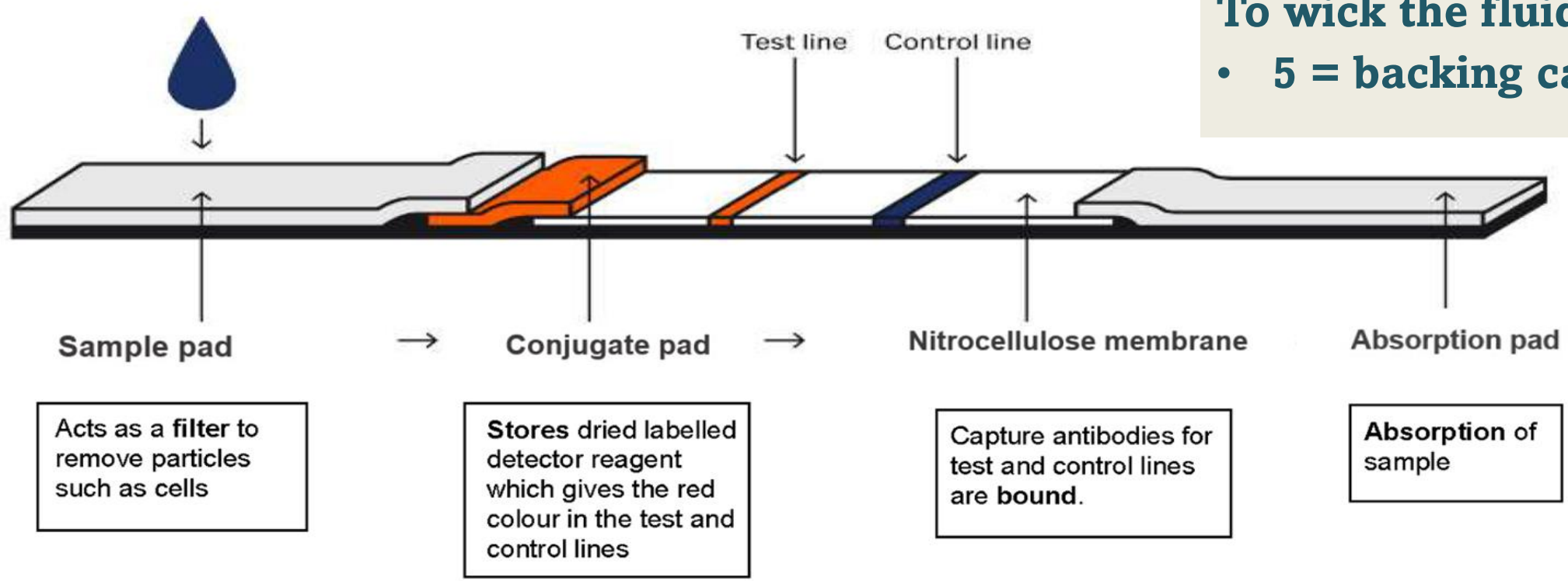
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# IMMUNOCHROMATOGRAPHIC METHOD: LATERAL FLOW STRIP TEST

## Test strip components



- Function of each components**
- 1 = sample application pad  
To drop sample and reduce non-specific binding of sample
  - 2 = gold nanoparticles conjugated pad  
To hold the detector particles
  - 3 = Analytical pad as detection zone  
To immobilize targeted antigen or antibody for analyzing
  - 4 = absorption pad  
To wick the fluid through the membrane and collect excess liquid
  - 5 = backing card for supporting strip components

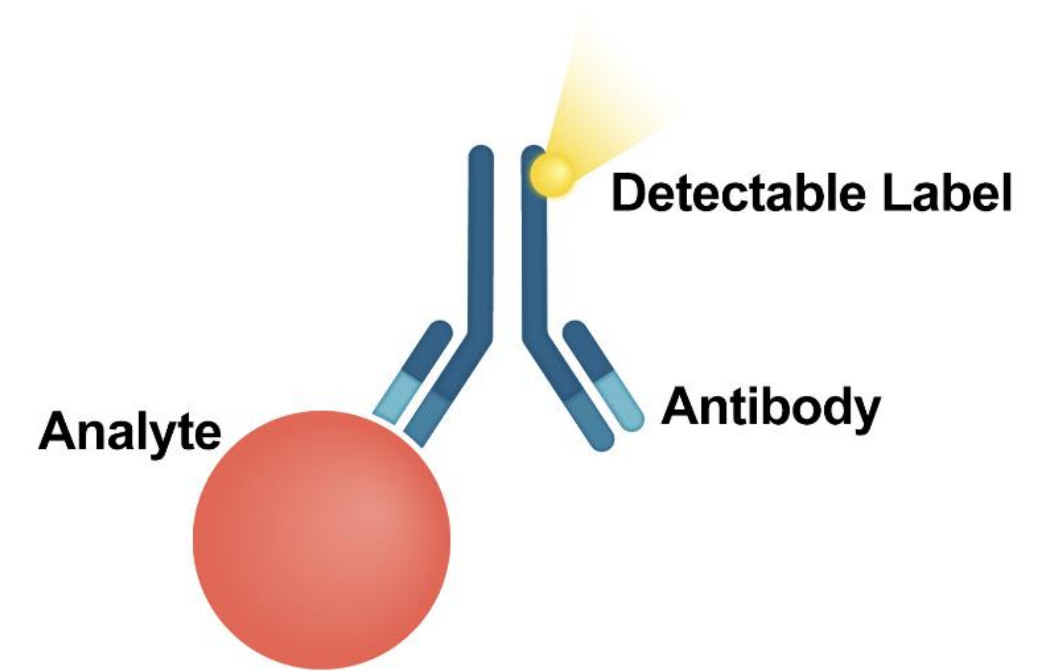


Picture from: <https://www.cytivalifesciences.com/en/us/news-center/lateral-flow-assay-format-sandwich-or-competitive-10001>



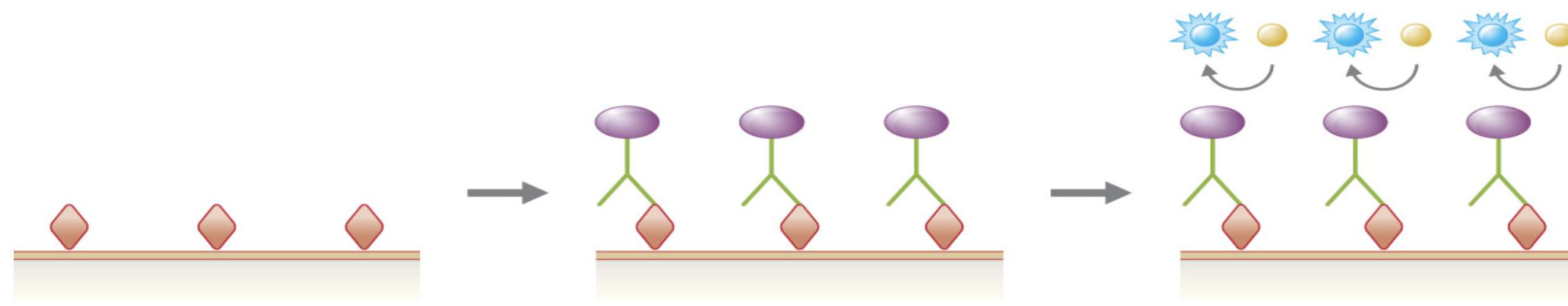
# Principle of immunoassay

- Immunoassay is a method for determining the content of a substance to be tested from a sample using the principle of immunology
- **This assay is a procedure for detecting or measuring specific protein or other substances through their properties as antigen and antibody.**
- Several methods are divided according to different labeling techniques: **fluorescent immunoassay**, **radioimmunoassay**, **enzyme immunoassay**, **colloidal gold immunoassay**, and **chemiluminescent immunoassay**

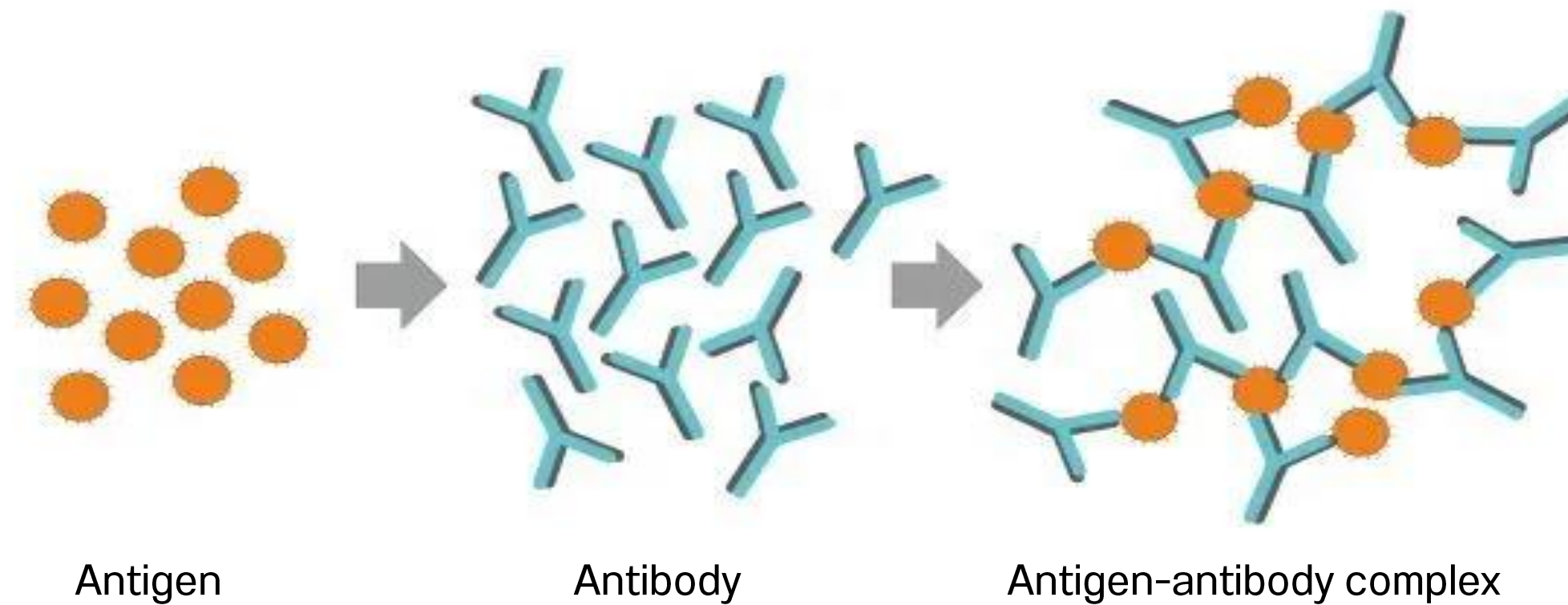


- Based on specific immuno-reaction between antibody (Ab) and antigen (Ag)

- The most widely used are enzyme-linked immunoassay (ELISA) and colloidal gold immunochromatographic strip test



# Antigen vs. Antibody



**An antigen** is any substance that stimulates an immune response.

- Foreign antigens come from outside the body and may be pathogens (like disease-causing viruses or bacteria) allergens (like pollen), or toxic substances (like venom or chemicals).
- Autoantigens originate in the body and do not usually provoke an immune response, except in individuals with autoimmune disorders.

**An antibody** is a Y-shaped protein that is produced by white blood cells, and that tags antigens for destruction by immune cells.

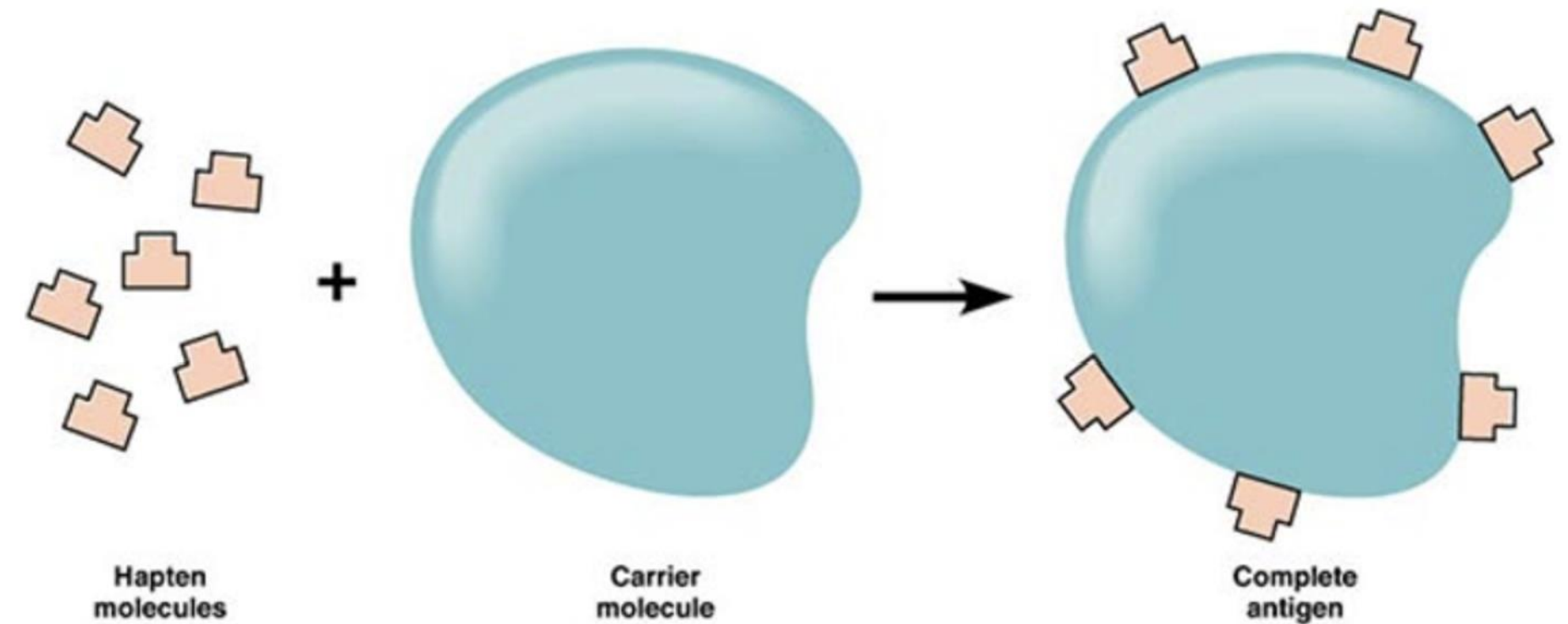
- When an antibody binds to an antigen, an antigen-antibody complex is formed.
- The formation of the antigen-antibody complex stimulates an immune response against the foreign substance.

Characteristics	Antigen	Antibody
Molecule Type	Usually, proteins may also be polysaccharides, lipids or nucleic acids.	Proteins
Definition	These are substances that provoke an immune response.	These are Glycoproteins that are secreted by immune cells (plasma cells) in response to a foreign substance (antigen).
Effect	Cause disease or allergic reactions.	Protect the system by lysis of antigenic material.
Origin	Within the body or externally.	Within the body.
Parts	Highly variable with different structural conformations and is usually composed of different epitopes.	Composed of three main parts: -Two light chains -Two heavy chains -Four polypeptides
Prevalence	Exists in all types of cells; mostly found in viruses, bacteria, and fungi.	Only present in some types of cells.
Synonyms	Immunogens	Immunoglobulins
Specific binding site	Epitope	Paratope
Complexity	Medium; exists due to random mutations in the cell's gene.	Very High; Complex chemical that bonds to a very specific Antigen.
Source	Usually from a foreign substance (viruses, and bacterial and fungal toxins).	Naturally produced by the body (B lymphocytes or B cells).
Kinds	There are three basic kinds of antigens (Exogenous, Endogenous, and Autoantigens)	There are five basic kinds of antibodies (IgG, IgM, IgA, IgE, and IgD).
Examples	<b>Exogenous antigens:</b> bacteria, viruses, fungi, etc. <b>Endogenous antigens:</b> Blood group antigens, HLA (Histocompatibility Leukocyte antigens), etc. <b>Autoantigens:</b> Nucleoproteins, Nucleic acids, etc.	Breast milk, tears, saliva, sweat, and mucus.

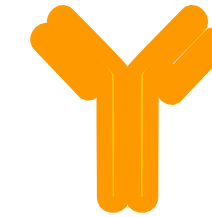
# Hapten

Hapten is a molecule that reacts with specific antibody but is not immunogenic by itself, it can be made immunogenic by conjugation to a suitable carrier.

Many drugs like penicillins are [haptens](#). A hapten is essentially an incomplete antigen. These small molecules can elicit an immune response only when attached to a large carrier such as a protein; the carrier typically does not elicit an immune response by itself.



# Monoclonal antibody VS Polyclonal antibody



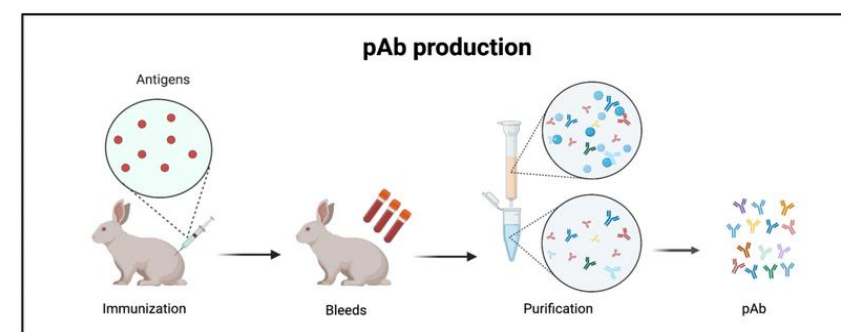
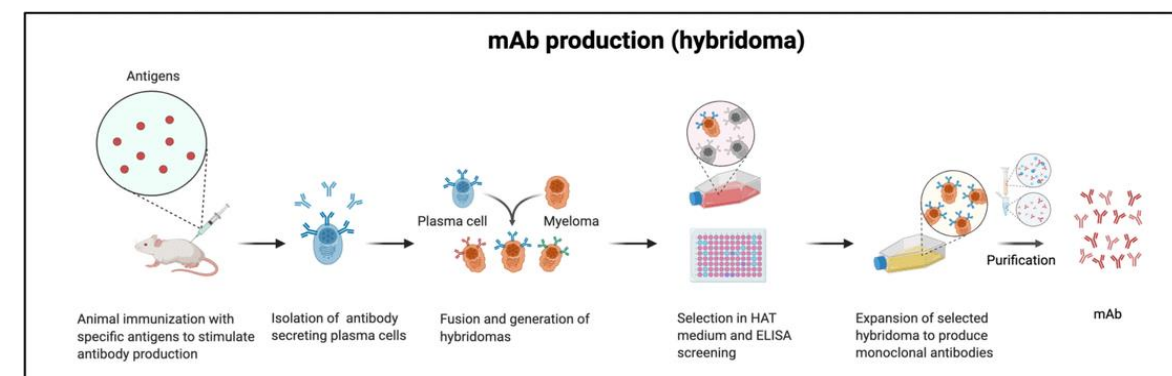
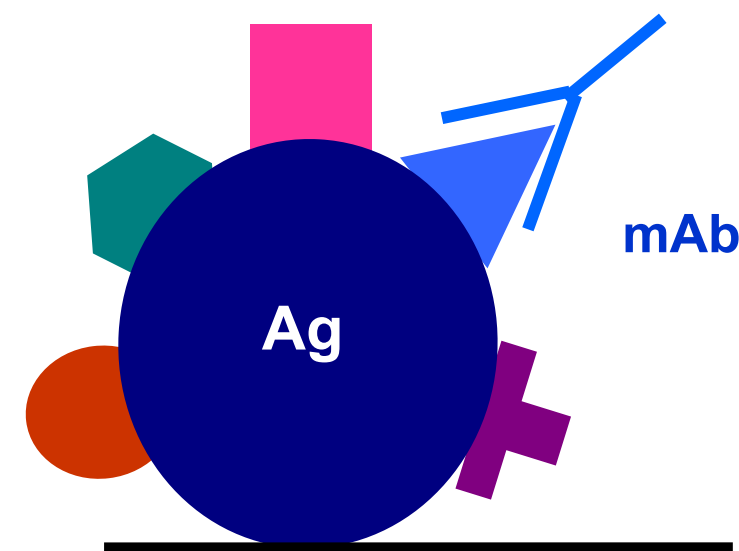
## Monoclonal antibodies (mAb)

- A homogenous antibody population.
- **Single antigenic determinant**
- Interact with a particular epitope on the antigen (mAb is antibody against only one epitope carried on antigen/immunogen)
- Refer to a homogenous population of antibodies that are produced by a single clone of plasma B cells.
- Produced by the same clone of plasma B cells.

## Polyclonal antibodies (pAb)

- A heterogeneous antibody population.
- Interact with different epitopes on the same antigen.
- Refer to a mixture of immunoglobulin molecules that are secreted against a particular antigen.
- Produced by different clones of plasma B cells.

**\*\*Epitope is the site of antigen which antibody come to attach.**

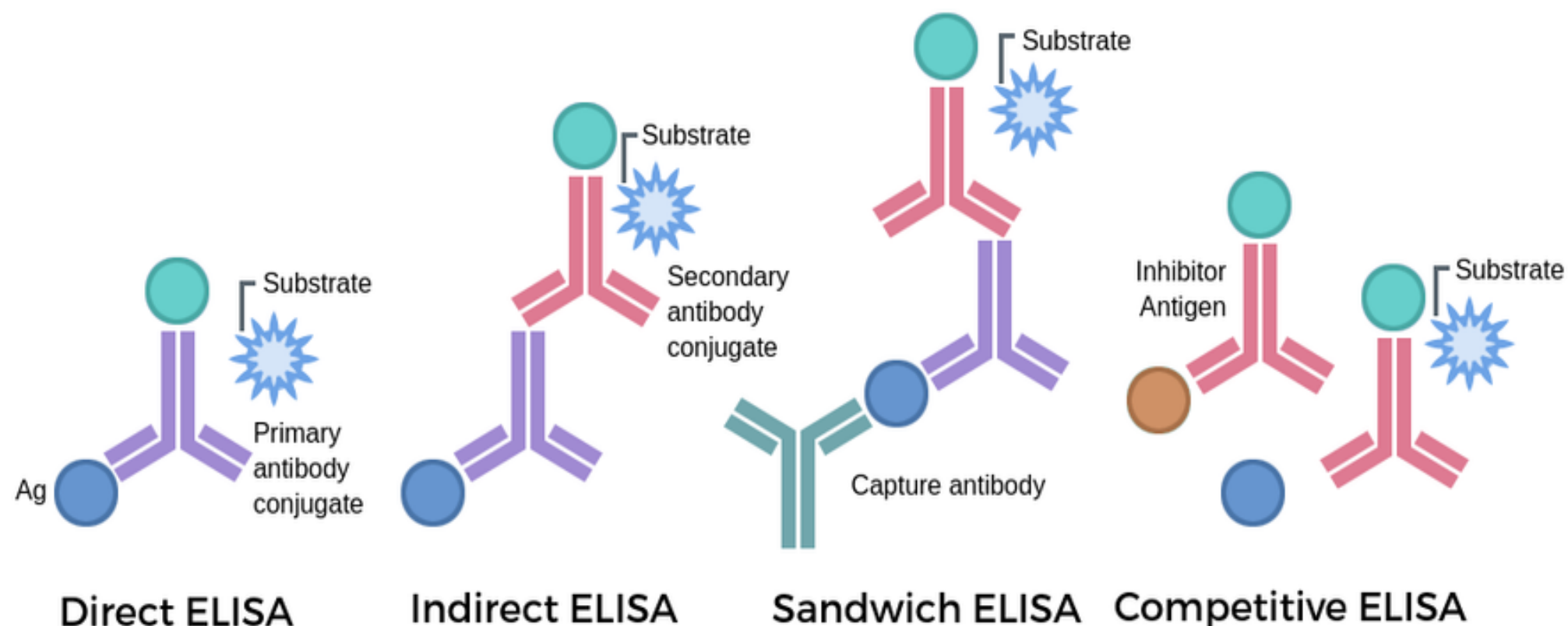


# Enzyme-Linked Immunosorbent Assay (ELISA)

ELISA (which stands for enzyme-linked immunosorbent assay) is a technique to detect the presence of antigens in biological samples. An ELISA, like other types of immunoassays, relies on antibodies to detect a target antigen using highly specific antibody-antigen interactions.

## There are four major types of ELISA:

1. Direct ELISA (screening antigen)
2. Indirect ELISA (antigen-coated plate; screening antigen/antibody)
3. Sandwich ELISA (antibody-coated plate; screening antigen)
4. Competitive ELISA (screening antibody)



## Diagnostic Tests

### Detect and Measure the Presence of Antibodies in the Blood

- Autoantibodies (anti-dsDNA, anti-dsg1, ANA, etc.)
- Antibodies against infectious disease (antibacterial, antiviral, antifungal)
- Hepatitis A, B, C, HIV, etc.

### Detect and Estimate the Levels of Tumor Markers

- Prostate-specific antigen (PSA)
- Carcinoembryonic Antigen (CEA)

### Detect and Estimate Hormone Levels

- Luteinizing hormone
- Follicular stimulating hormone
- Prolactin
- Testosterone
- Human chorionic gonadotropin (hCG)

### Tracking Disease Outbreaks

- Cholera
- HIV
- Influenza
- Covid-19

### Detecting Past Exposures

- HIV
- Lyme disease
- Hepatitis

### Screening Donated Blood for Possible Viral Contaminants

- Anti-HIV-1/2
- Anti-HCV
- HBsAg

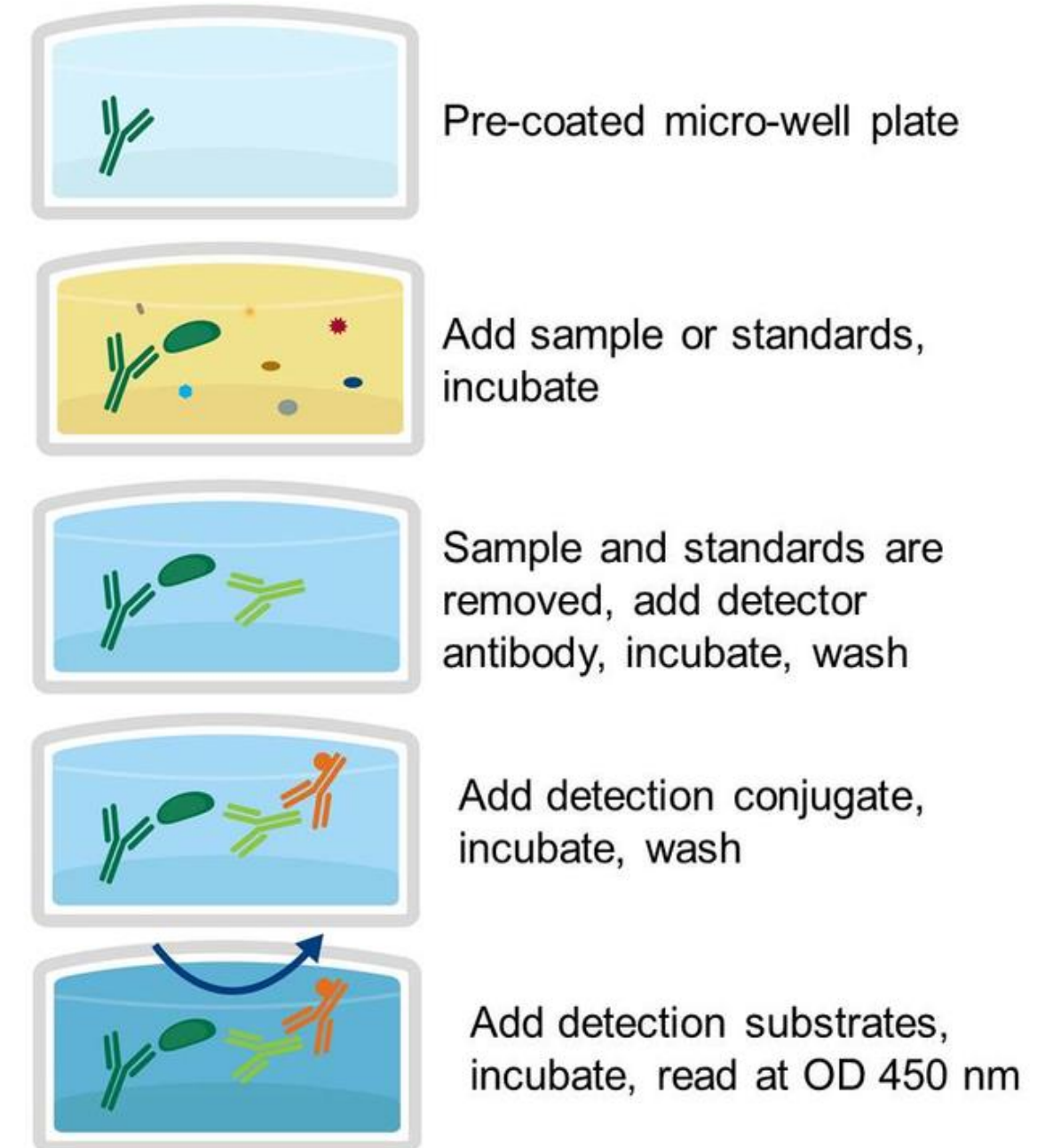
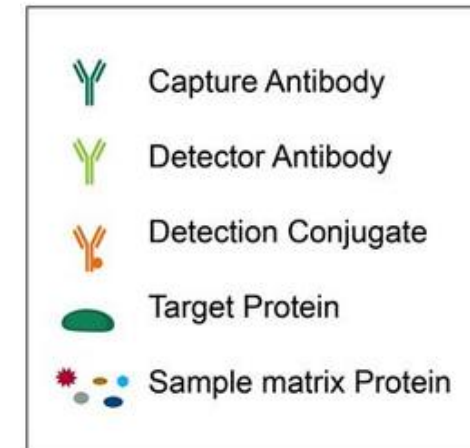
### Detecting Drug Abuse

- Amphetamine
- Methamphetamine
- 3,4-methylenedioxymethamphetamine
- Cocaine
- Benzoylcegonine

# Enzyme-Linked Immunosorbent Assay (ELISA)

There are four main general steps to completing an ELISA immunoassay. These steps are:

1. Coating (with either antigen or antibody)
2. Blocking (typically with the addition of bovine serum albumin [BSA])
3. Detection
4. Final read



Uses enzyme-antibody conjugates to quantify target molecules

Direct ELISA: Uses a single antibody to detect the presence of an antigen

Detection of HIV antigen p24 up to one month after being infected

Indirect ELISA: Measures the amount of antibody produced against an antigen

Detection of HIV antibodies in serum

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- A paper-based platform to detect analyte in sample
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- Capillary force moving
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- Getting results in 10-30 mins



- Can be applied for:-
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  - : Filed testing

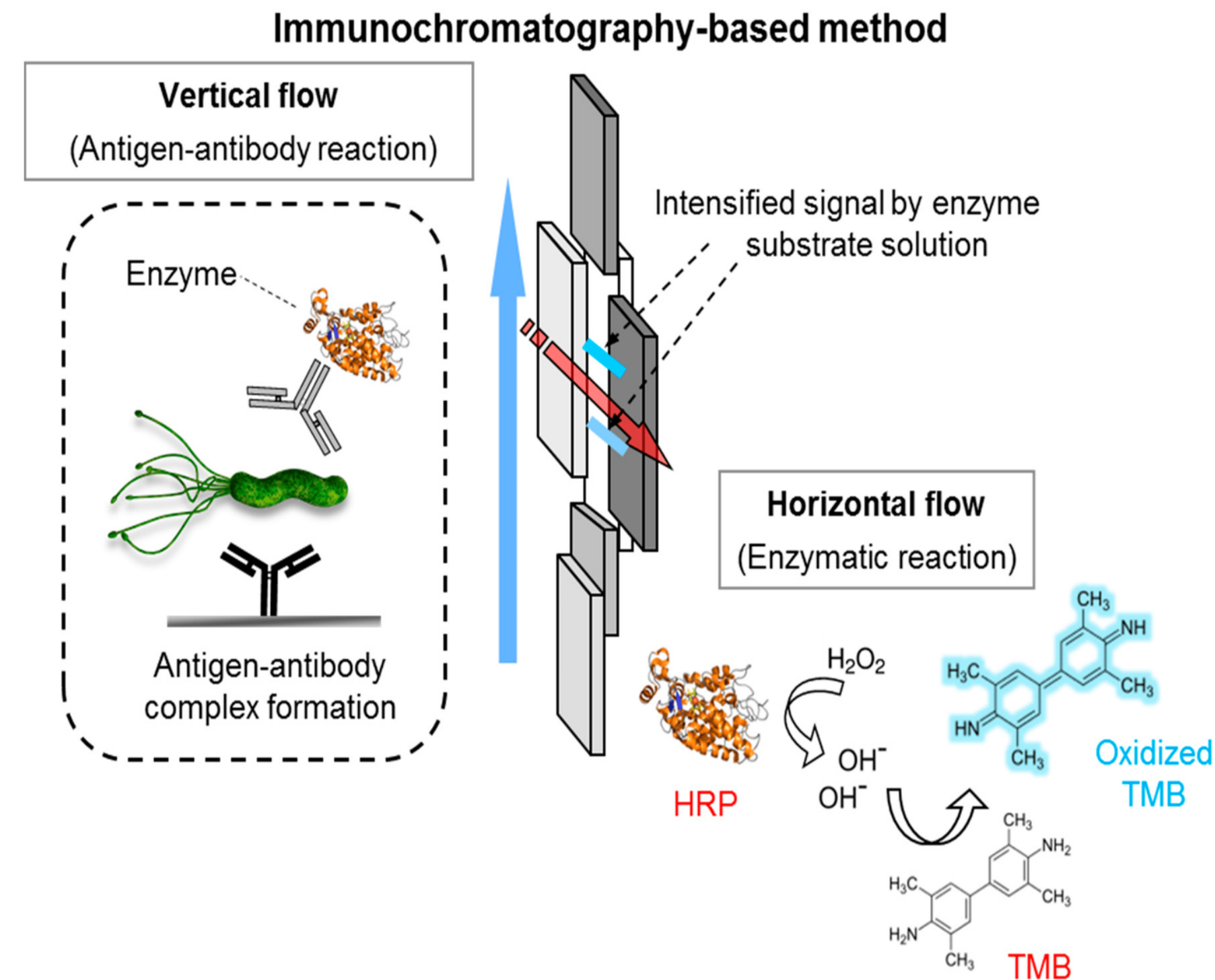
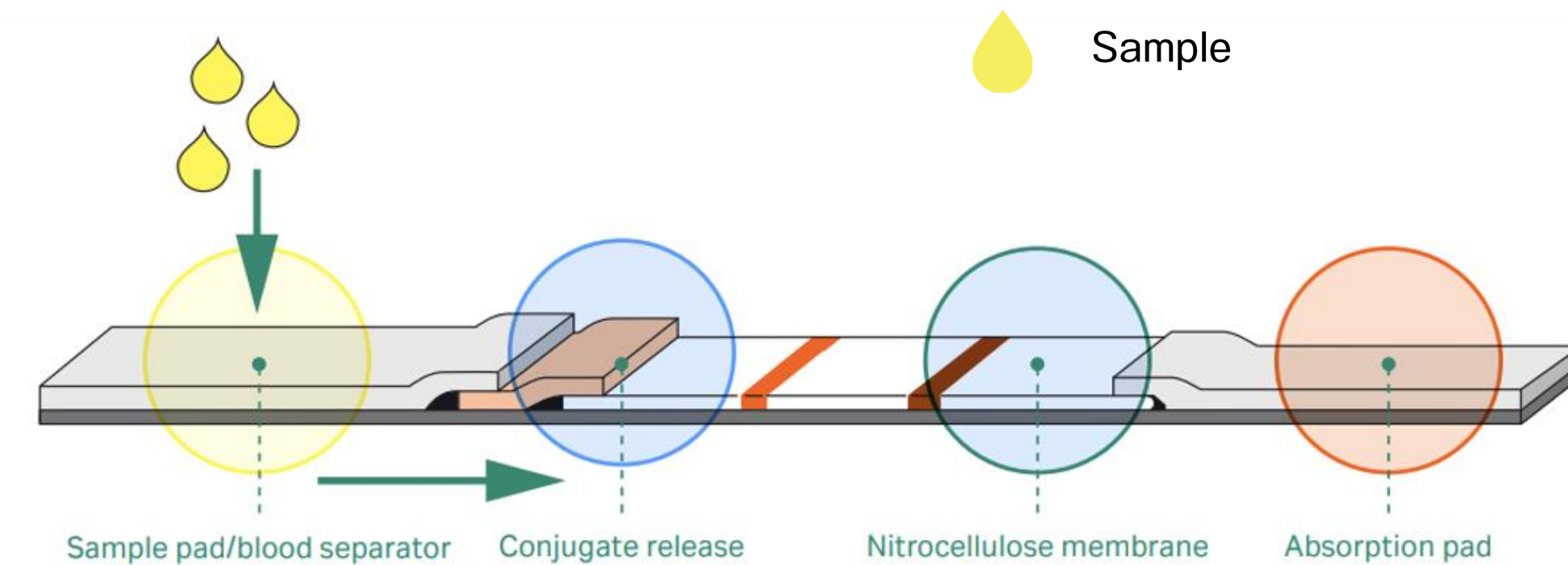


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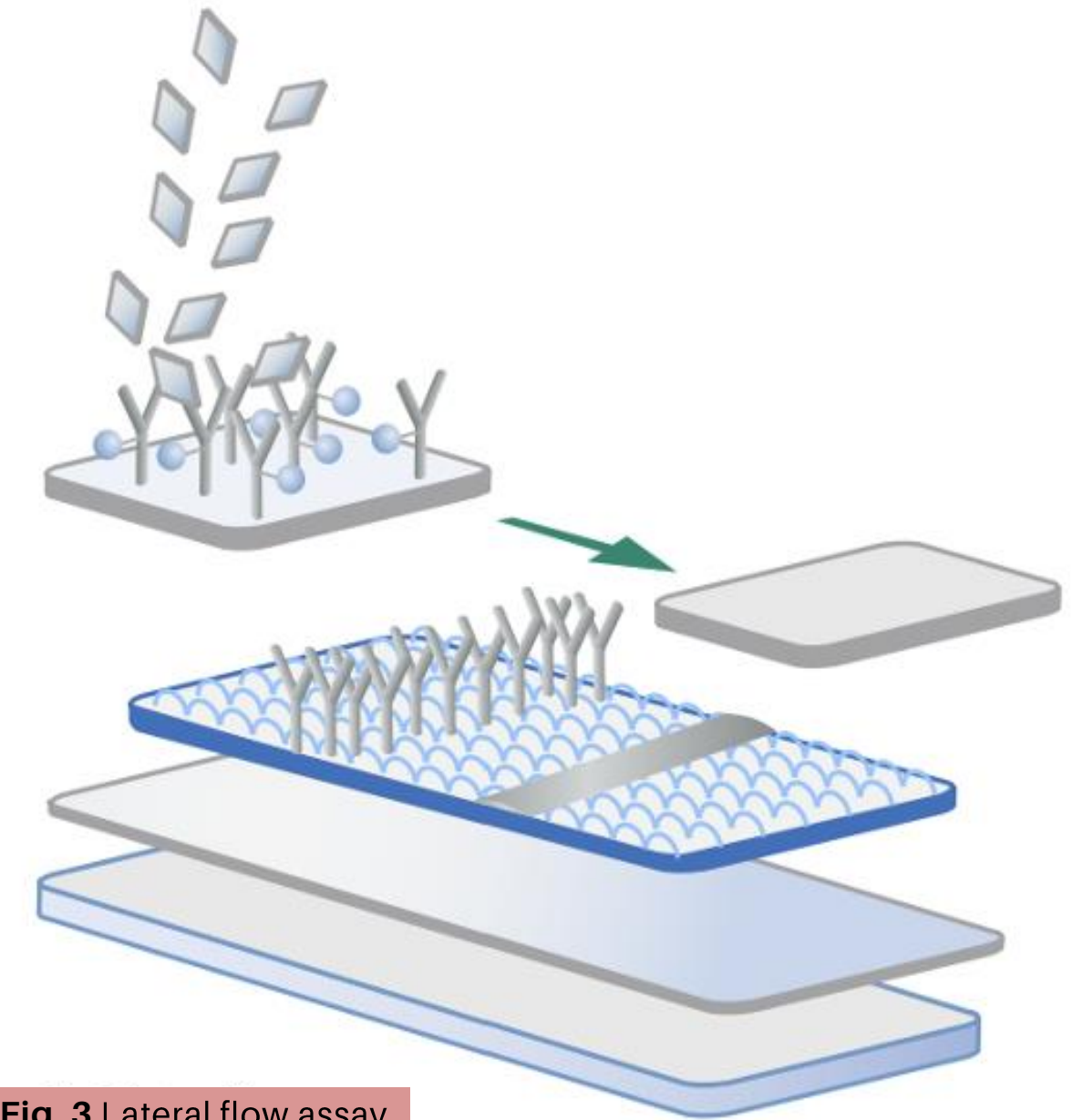


# IMMUNOCHROMATOGRAPHIC METHOD: LATERAL FLOW IMMUNOASSAY

The sample flows through the device and comes in contact with immobilized reagents. The antibody and analyte migrate to a capture zone of membrane-immobilized antibody. Any unreacted tagged antibody flows past the capture zone to absorbent.



**Fig. 2** Drawing of a strip test immunoassay



**Fig. 3** Lateral flow assay

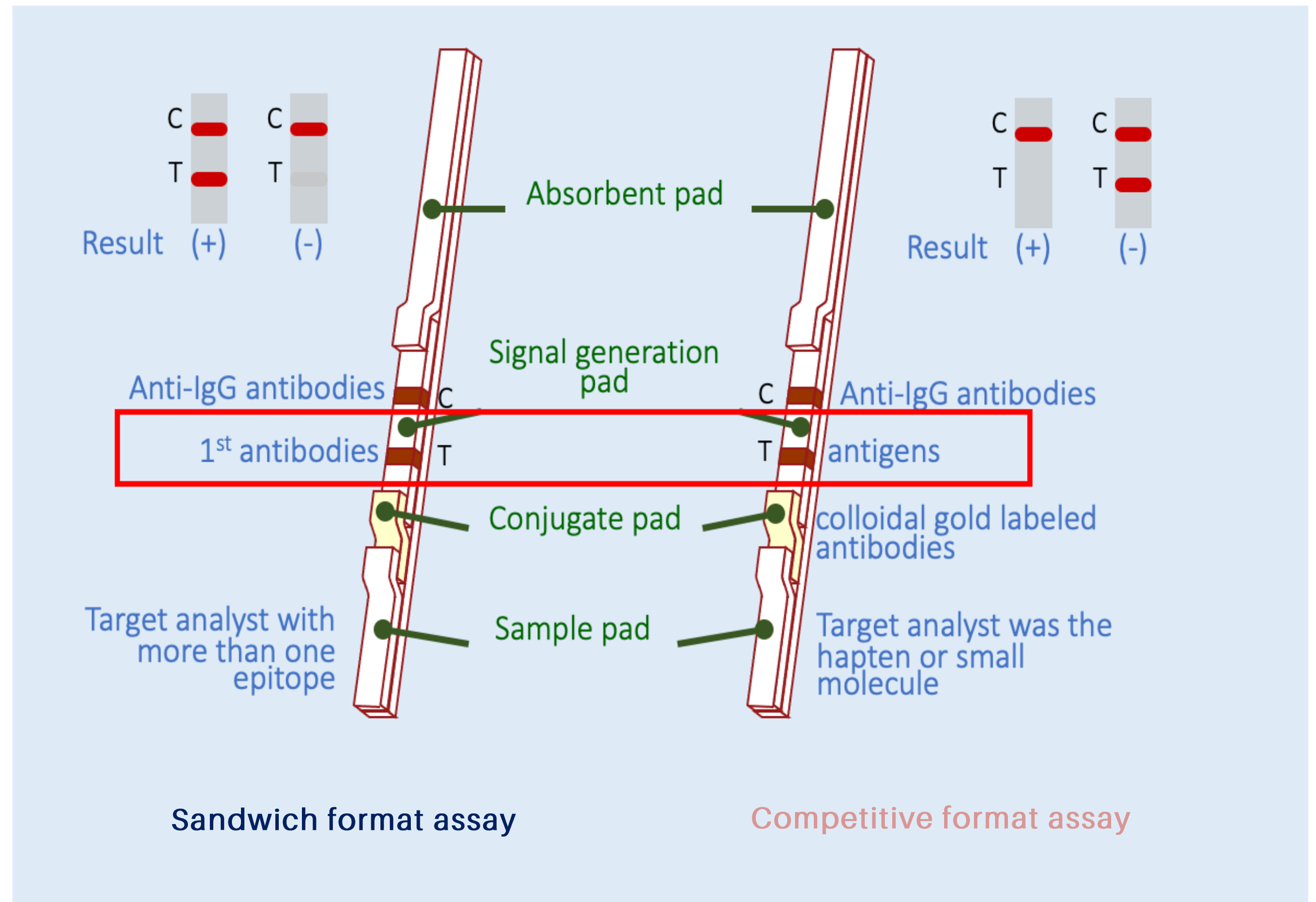
# TYPES OF IMMUNOCHROMATOGRAPHIC METHOD

## 1. Direct format (Non-competitive/Sandwich assay)

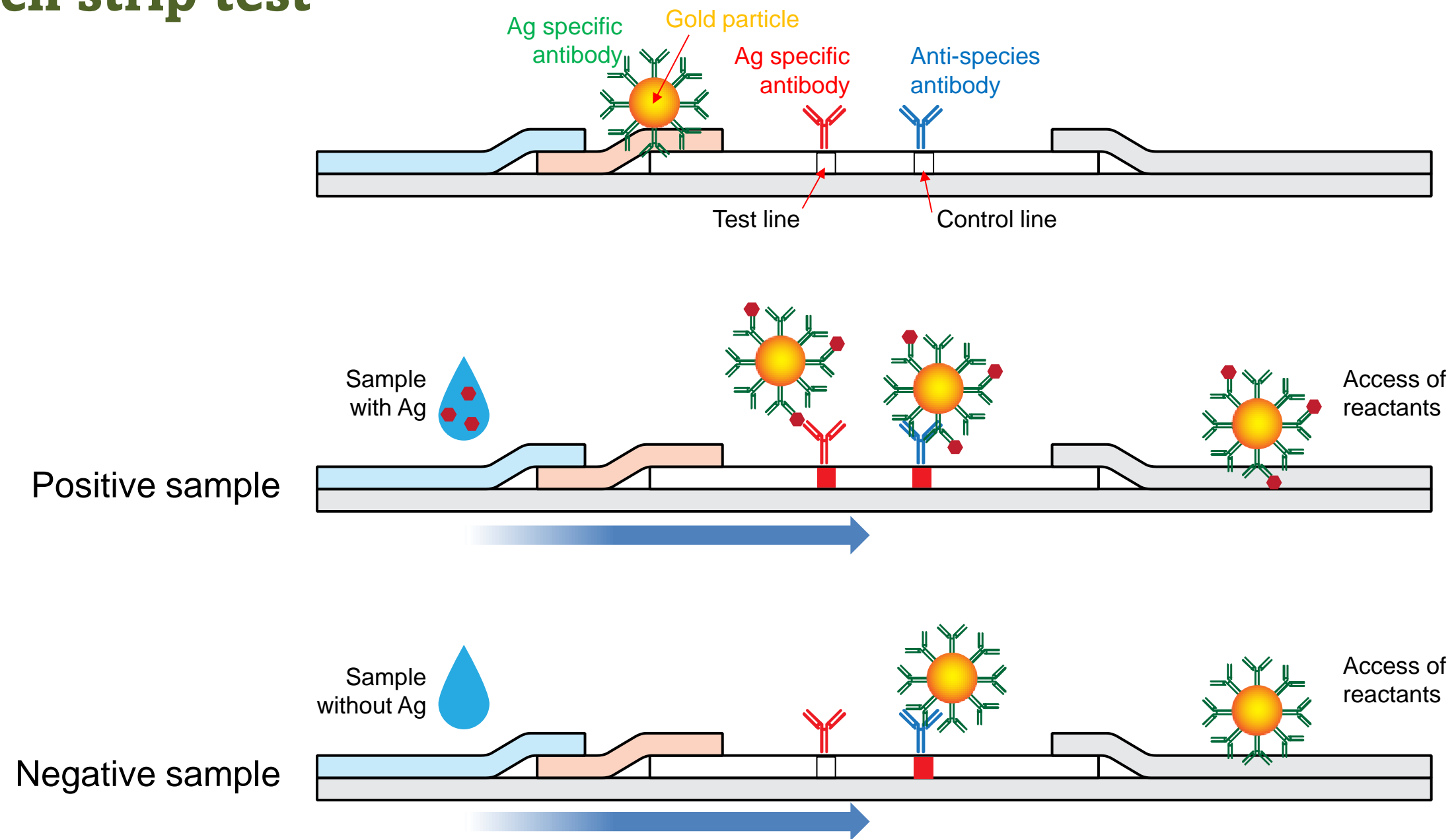
- Large analyze with several antigenic sites

## 2. Competitive (Competitive inhibition)

- Small molecule with a single antigen



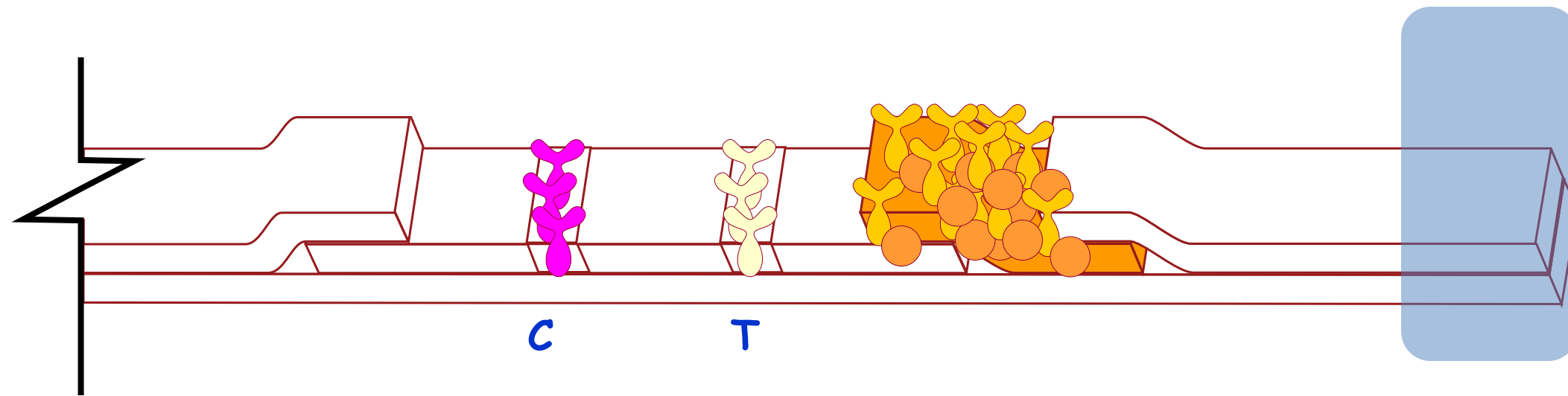
## Sandwich strip test



## 1. Direct format (Non-competitive/Sandwich assay)

### Test strip: Sandwich format assay

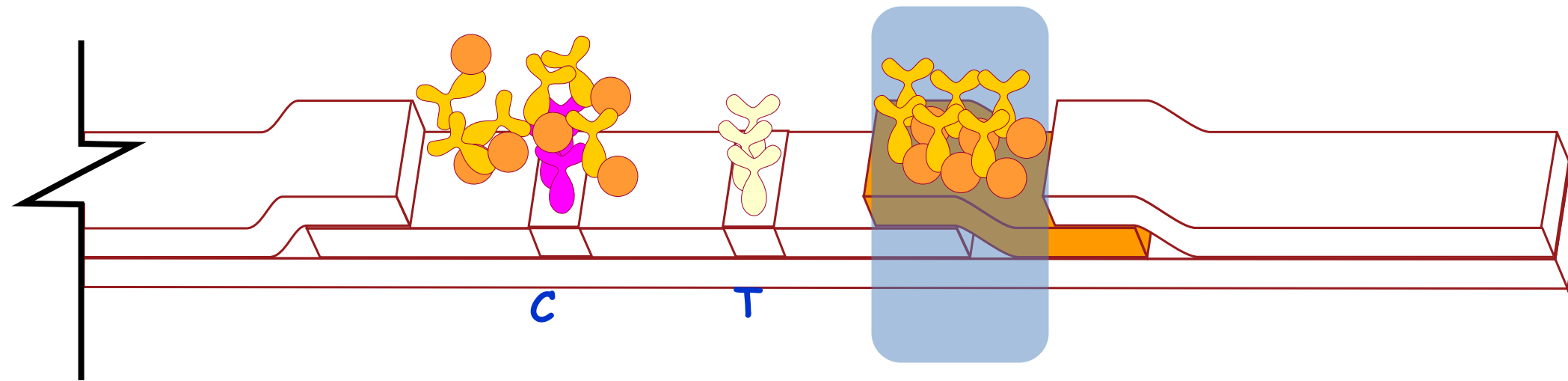
- Sample without antigen



# 1. Direct format (Non-competitive/Sandwich assay)

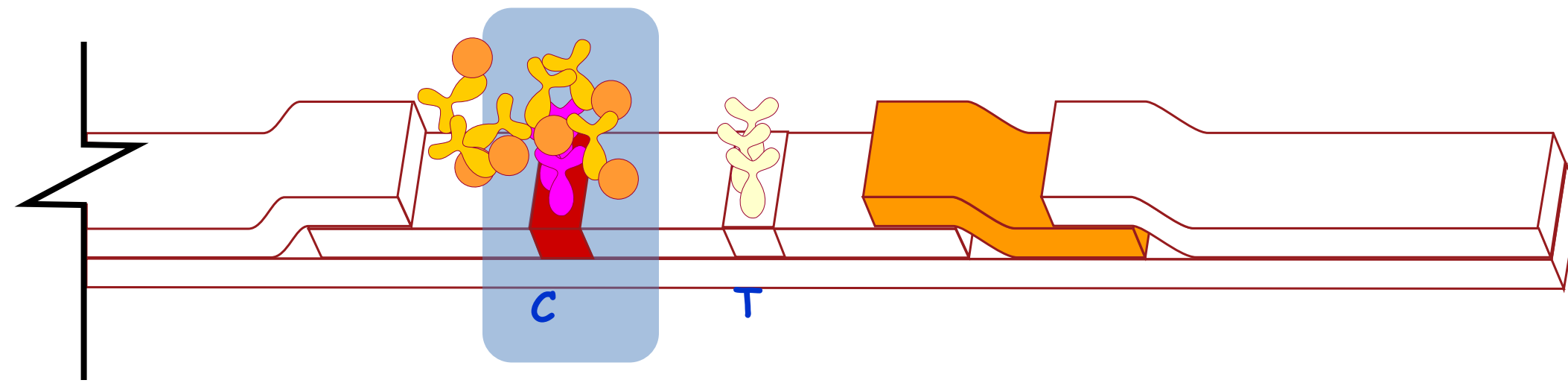
### Test strip: Sandwich format assay

- Sample without antigen



# 1. Direct format (Non-competitive/Sandwich assay)

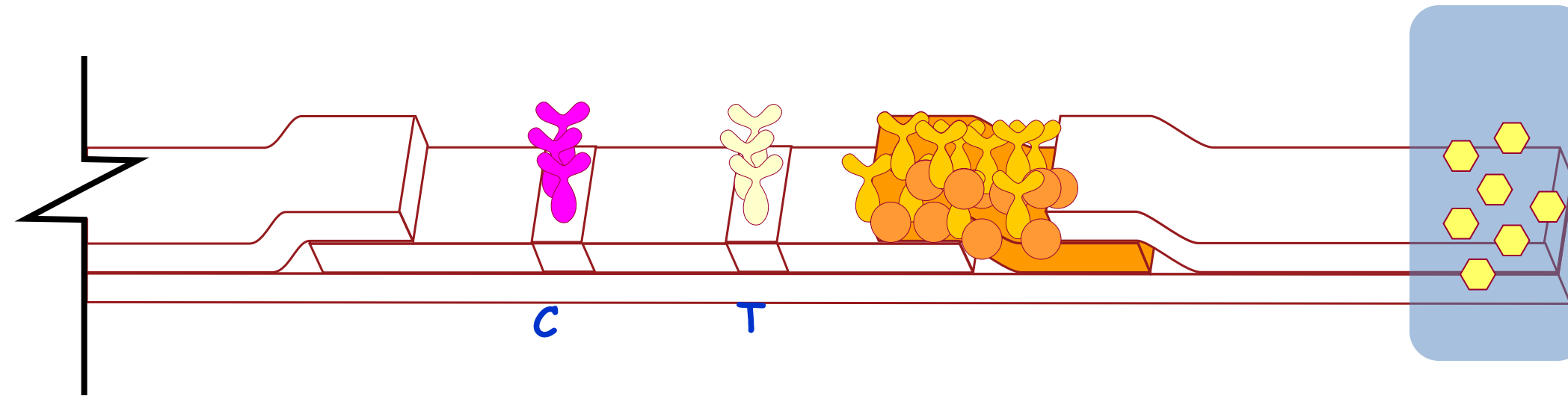
- Sample without antigen



Negative result

# 1. Direct format (Non-competitive/Sandwich assay)

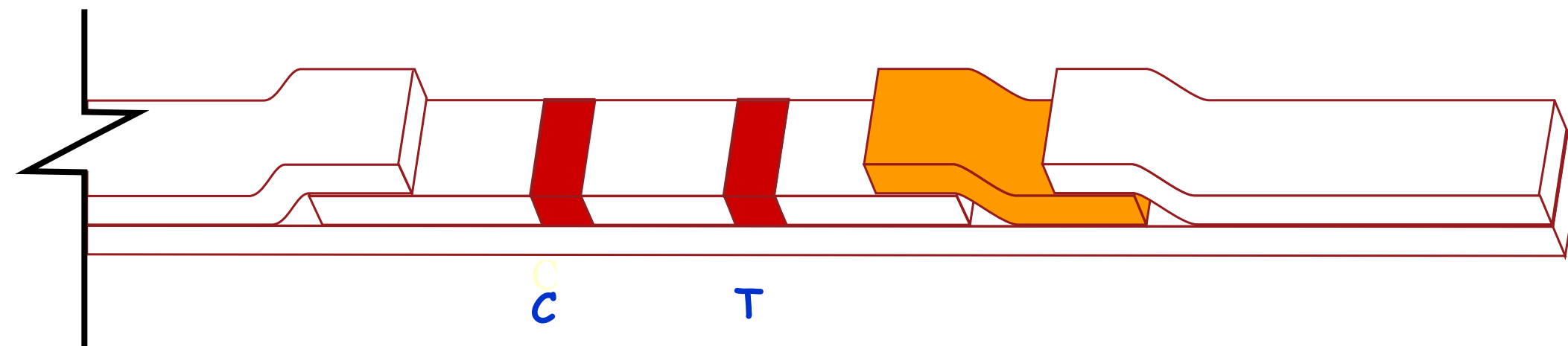
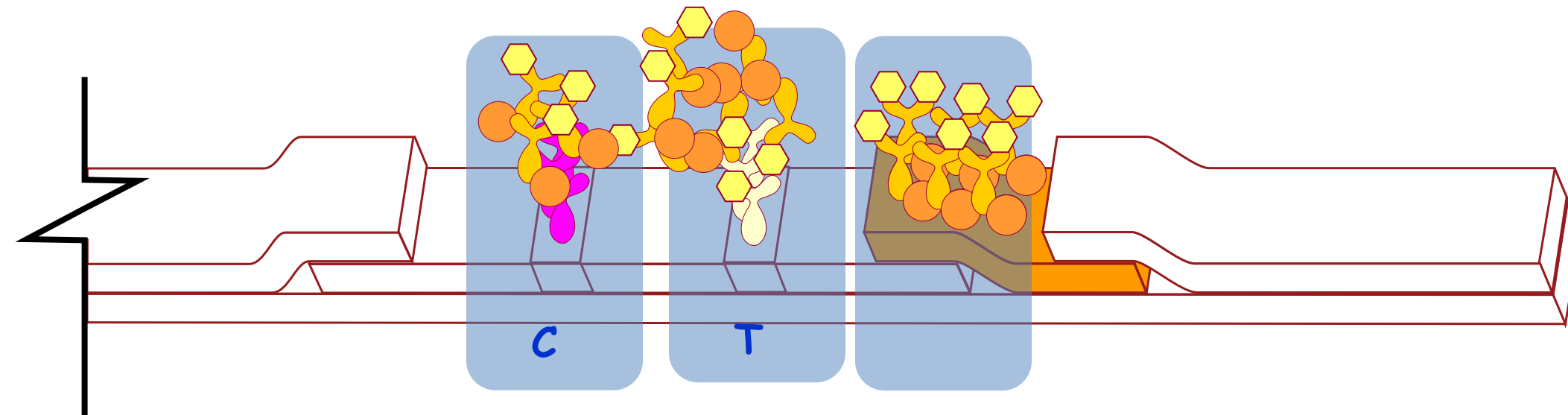
- Sample with antigen



# 1. Direct format (Non-competitive/Sandwich assay)

Test strip: Sandwich format assay

- Sample with antigen

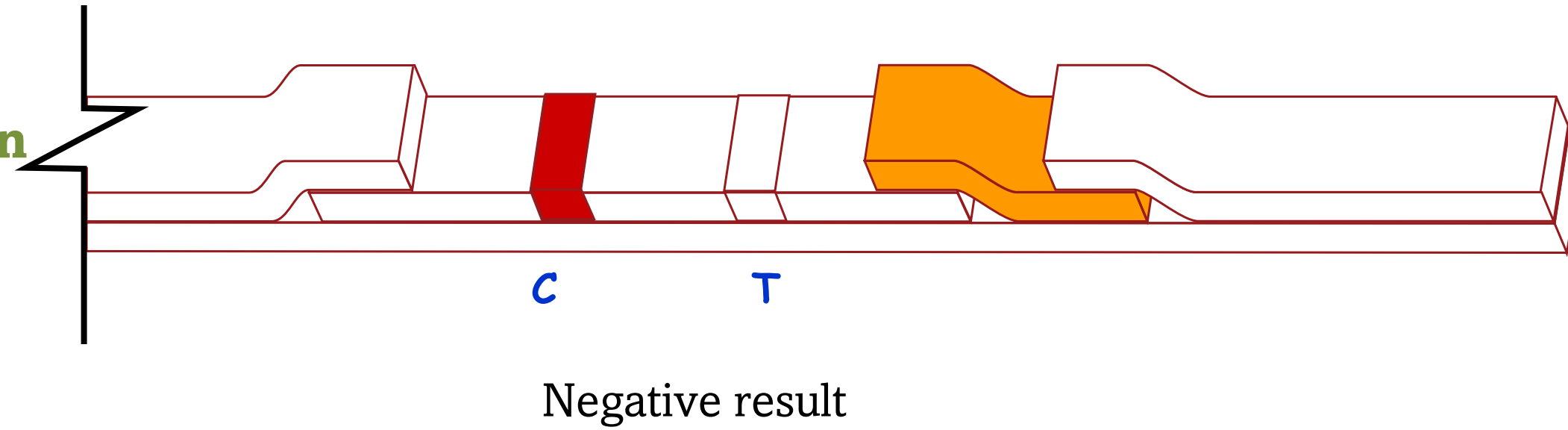


Positive result

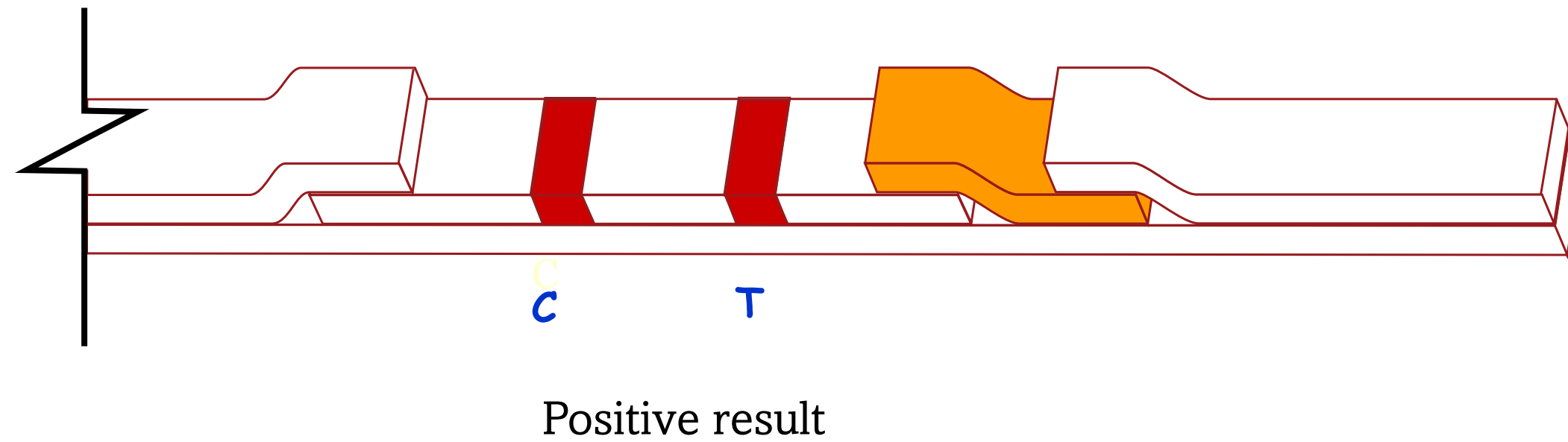


# 1. Direct format (Non-competitive/Sandwich assay)

- Sample without antigen



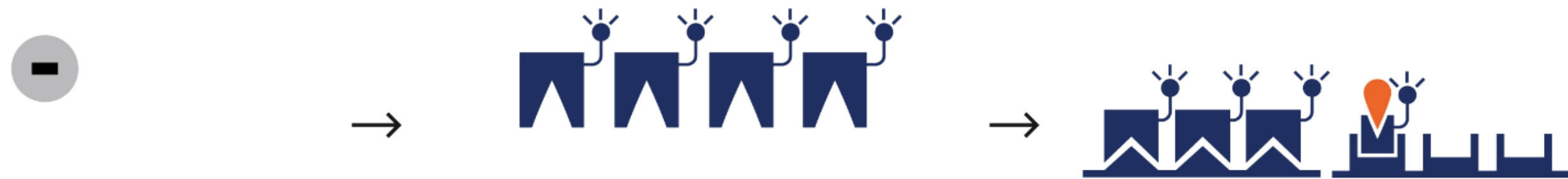
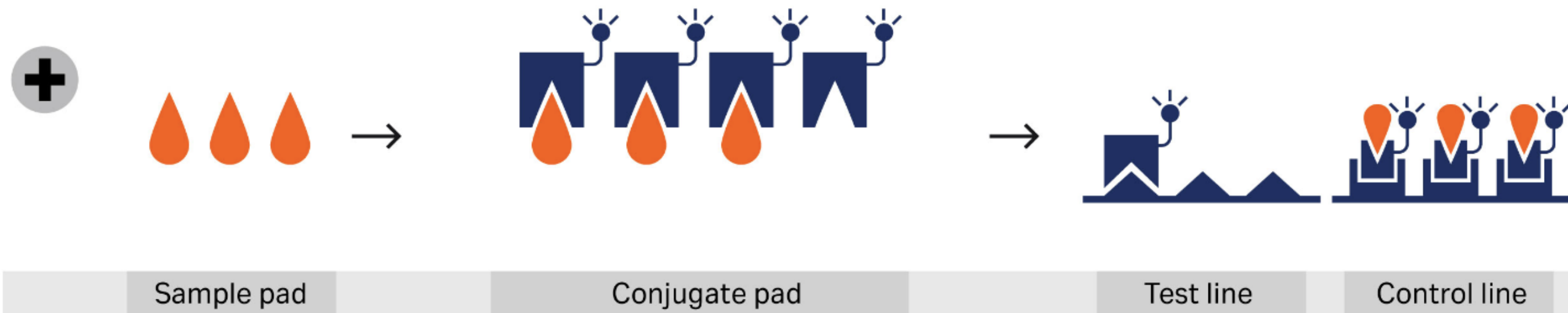
- Sample with antigen



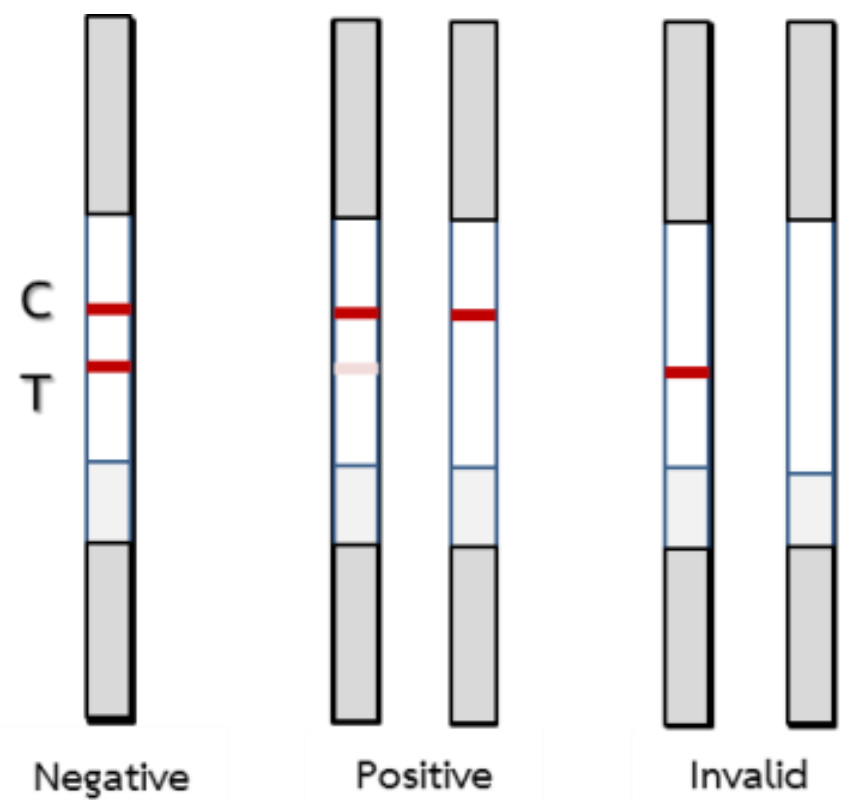
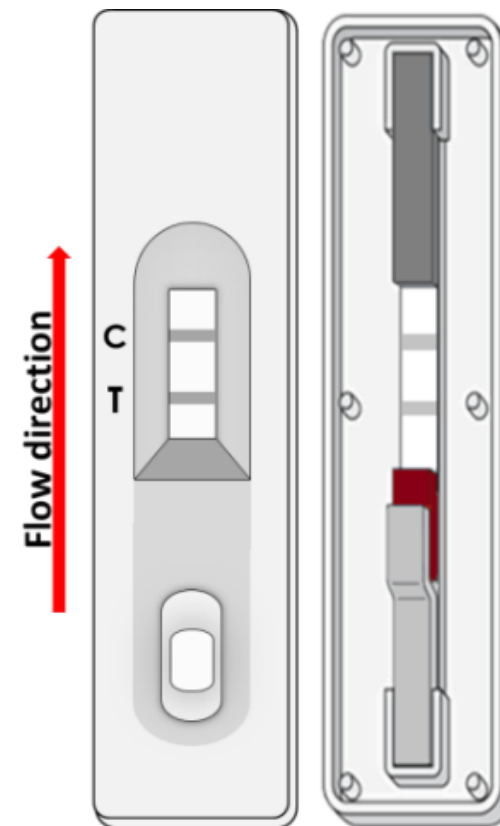
# Competitive Strip test (Competitive inhibition)

# Methods

## Competitive

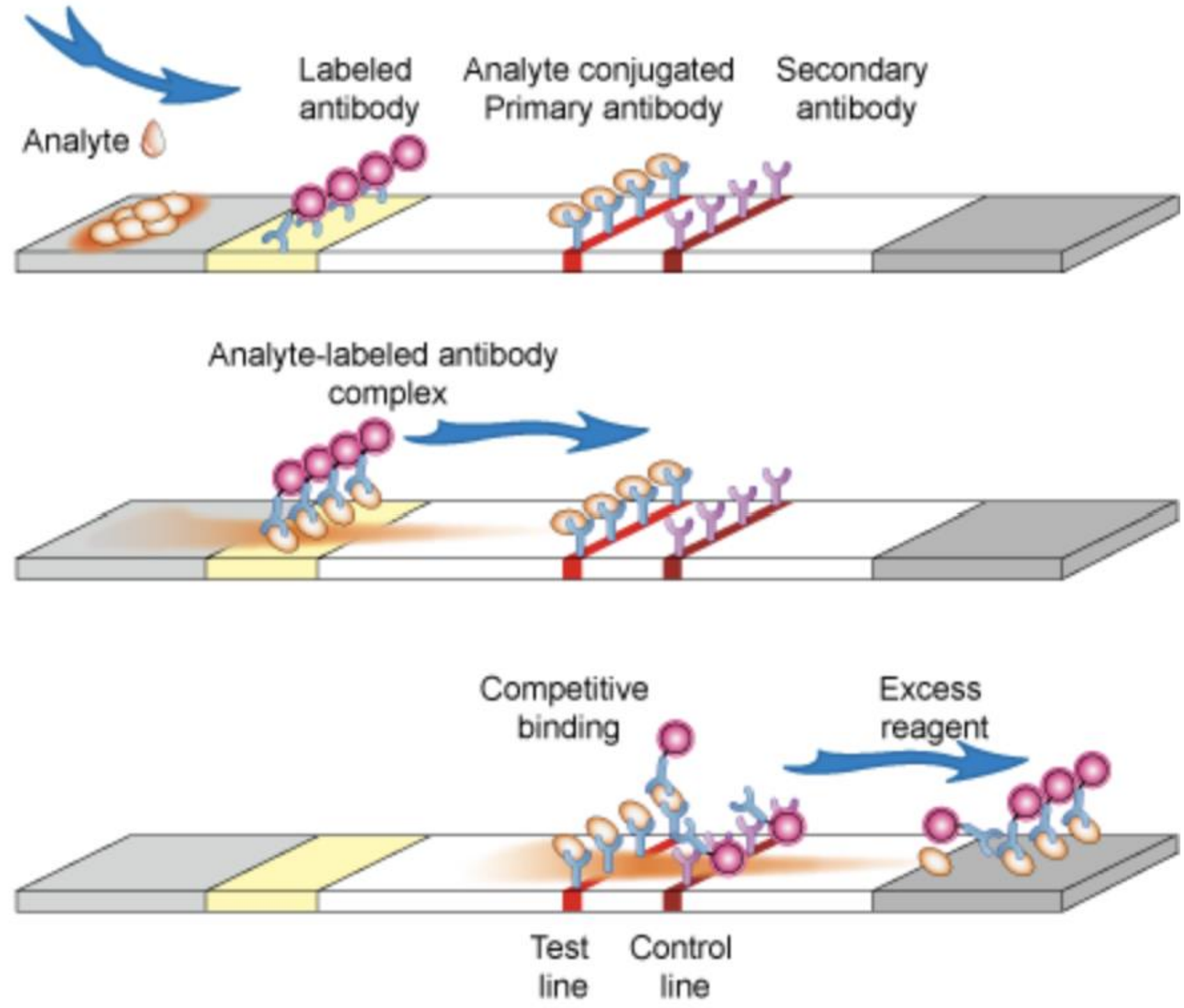


Target      Label-conjugate antibody      Imm. captured antibody



# Competitive Strip test (Competitive inhibition)

# Methods

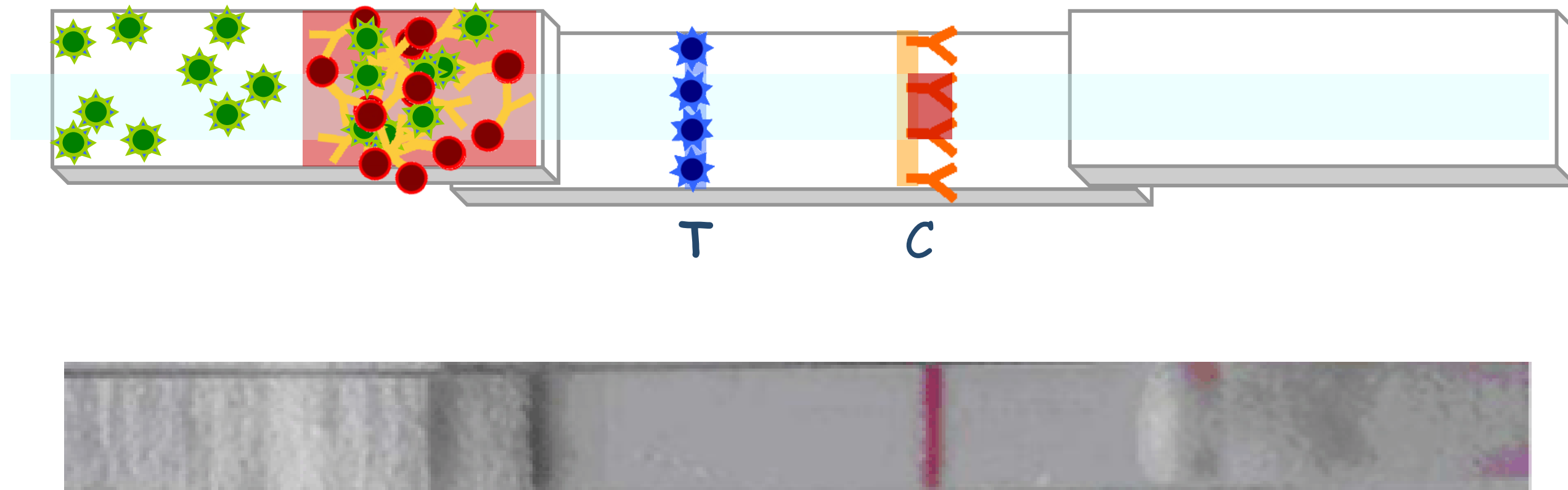


# Methods

## Test strip: Competitive format assay

- Sample with antigen

10 min

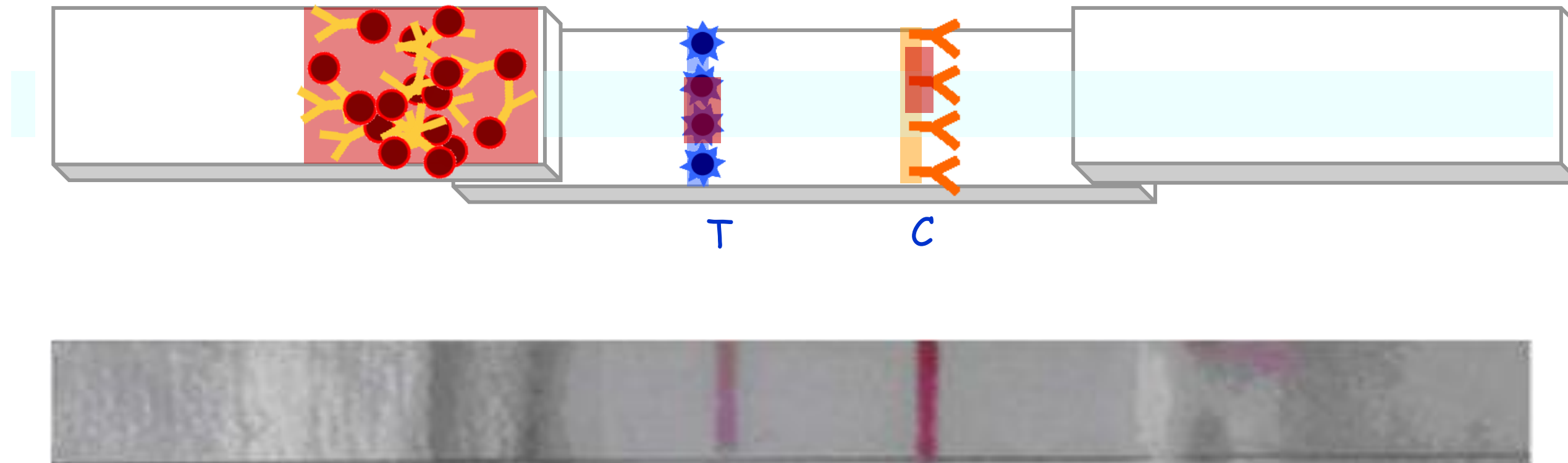


**Positive reaction**

# Methods

- Sample without Antigen

10 min

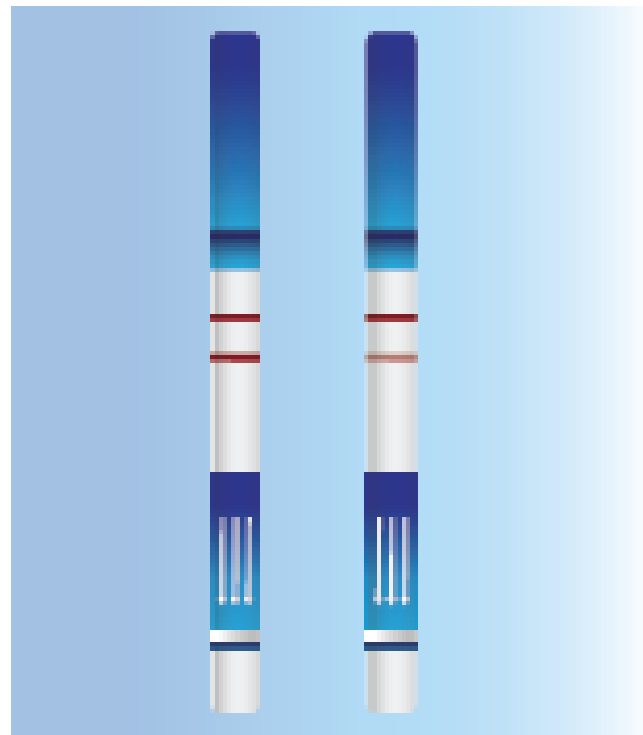


**Negative reaction**

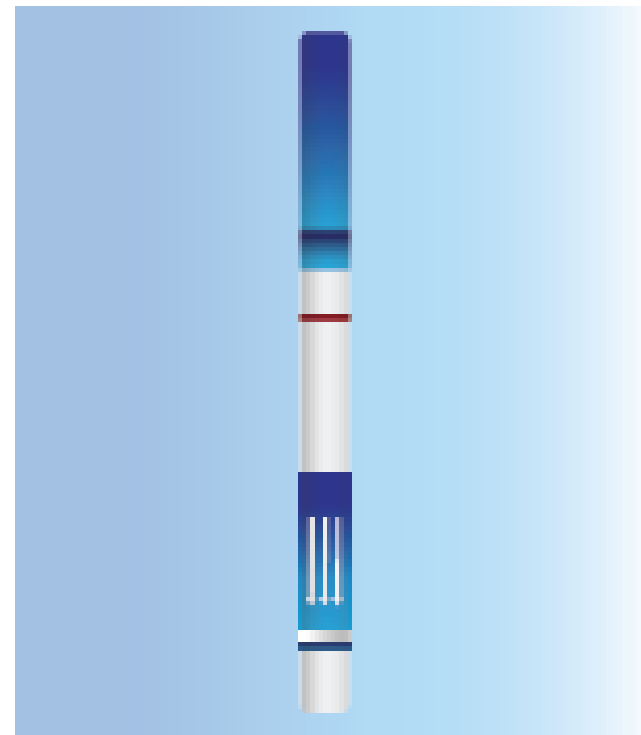
# How different of the result between Direct and Indirect assay

## 1. Direct format (Non-competitive/Sandwich assay)

- Large analyze with several antigenic sites



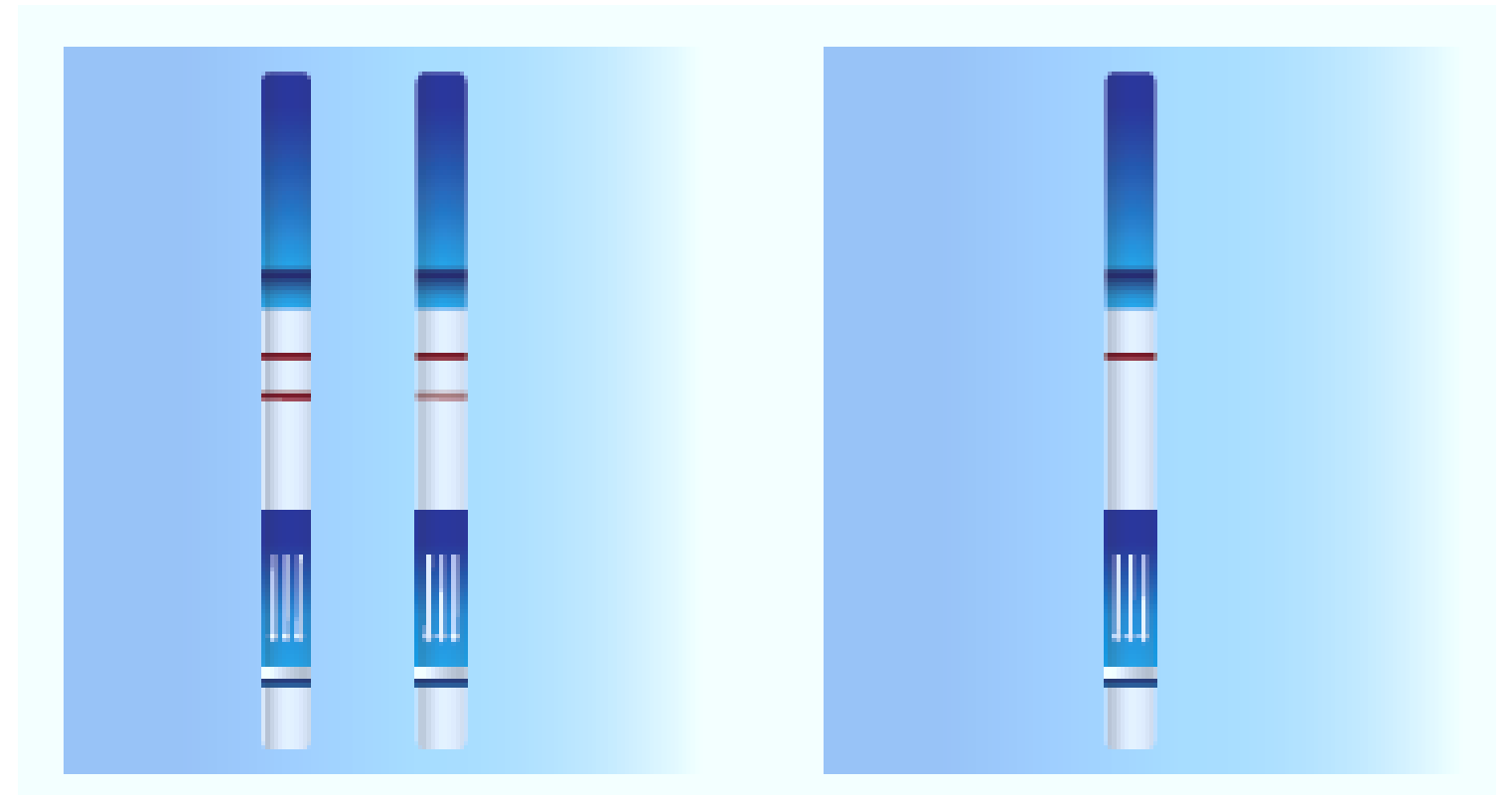
Positive result



Negative result

## 2. Competitive (Competitive inhibition)

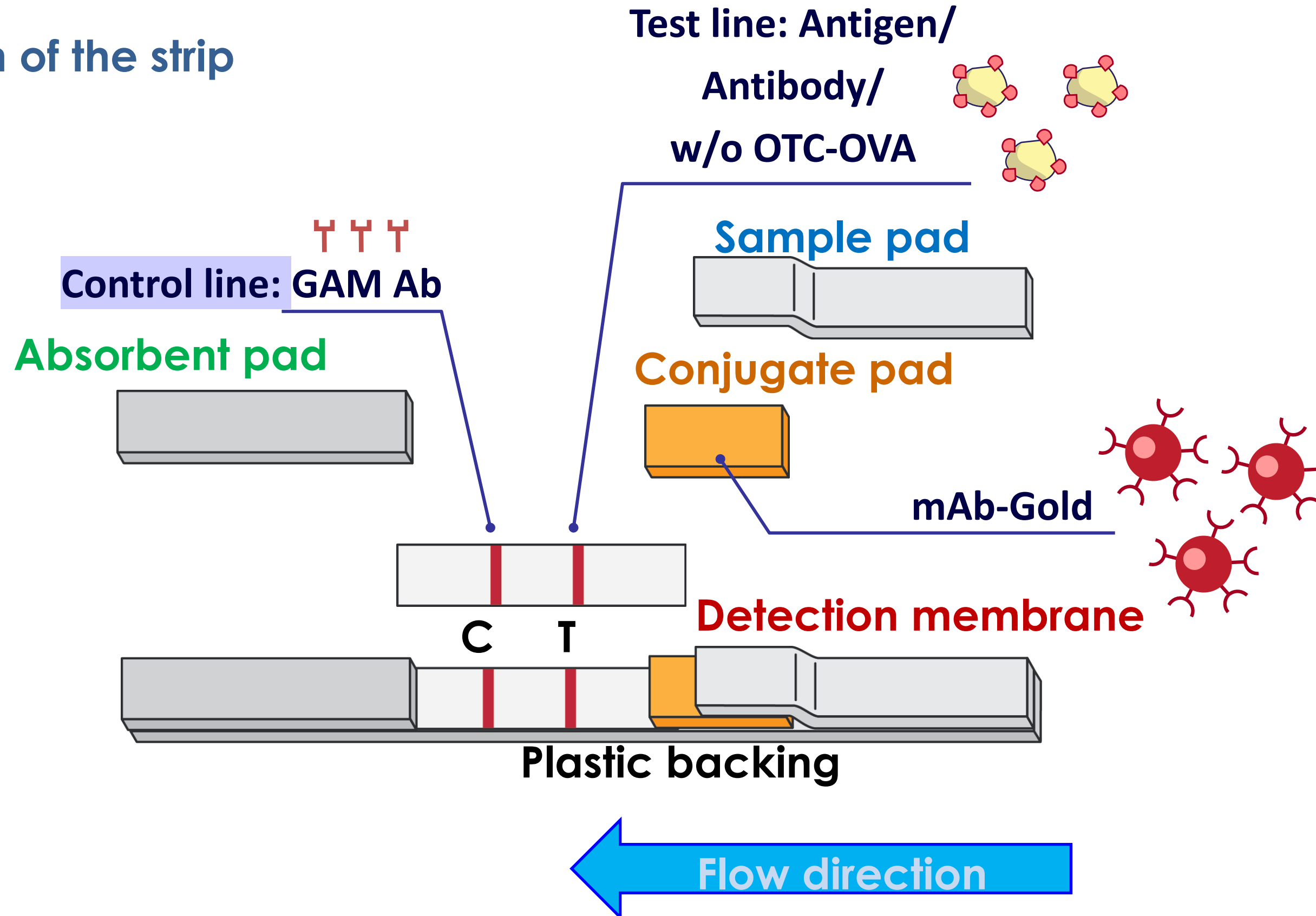
- Small molecule with a single antigen



Negative result

Positive result

- Composition of the strip



# DESIGN AND DEVELOPMENT

## Target product profile

## Design of the device

### 1 To define characteristic and acceptable criteria

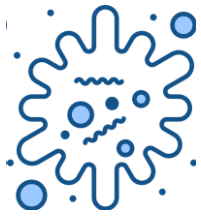
### 2 To define type of device

- Intended use
- Target analyte
- Specificity, sensitivity, and accuracy
- Sample type
- Time to get the result
- Type of analysis (qualitative/quantitative)
- Limit of detection/cut-off value
- Target/patient population

- Analyte Sample
- Material components
- Compatibility
- Recognition molecule such as antibody, aptamer
- Detection method

Intended use:

Analyte Sample:



Diagnosis

Veterinary

Agriculture

Food

Antigen

Antibodies

Nucleic Acids

Drug

Vitamins



# To Develop A Rapid Diagnostic Kit Using Immunochromatographic Method

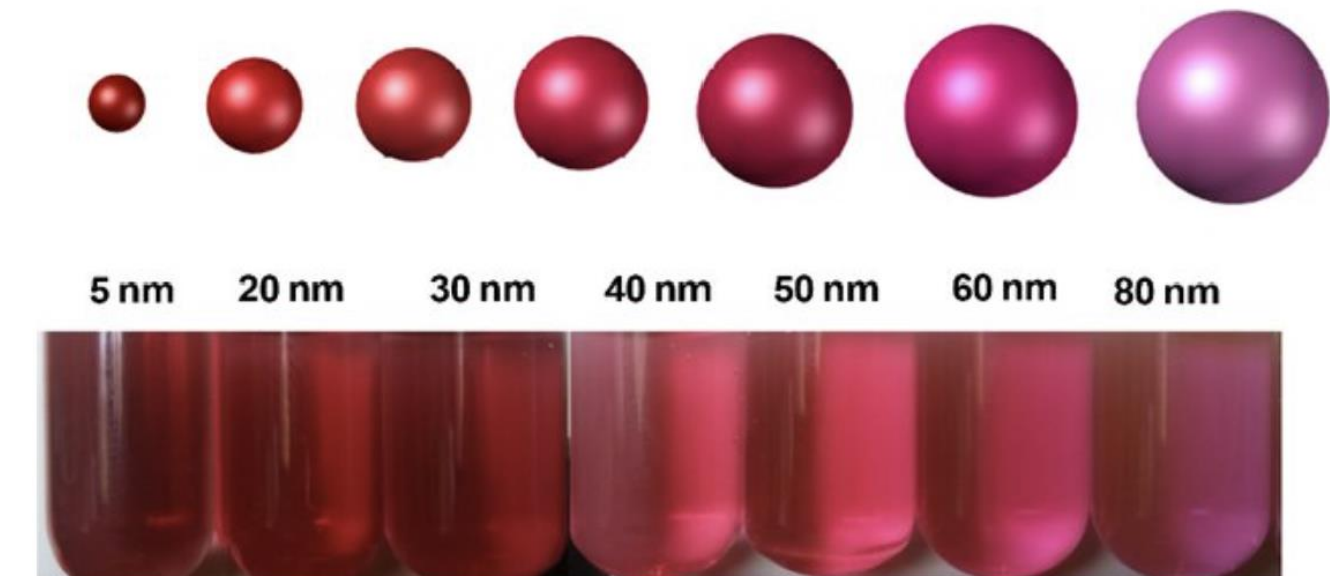
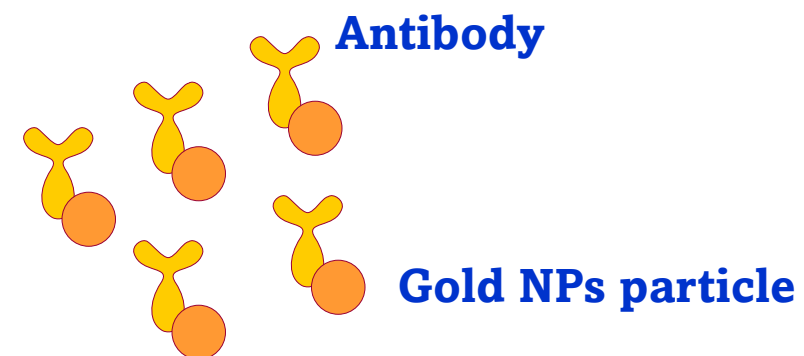


1. Production of antibody against antigen >> For detection
  - Immunization
2. Synthesis of colloidal gold >> For signal reporter
  - Tag-labeled antibody probe (Nanogold colloidal particle)
3. Formation of antibody-colloidal gold conjugates
  - Optimization antibody concentration to attach colloidal gold
4. Preparation of immunochromatography test strip
  - Optimization of antigen and antibody concentration to code on the analytic pad
5. Prototype testing
  - Process verification (partial method validation)
6. Performance testing (Method validation)
  - Performance testing (for finding: accuracy, specificity, sensitivity, false positive rate, and false negative rate)
  - Cross-reactivity testing
  - Stability testing

# • Synthesis of colloidal gold

## Colloidal gold probe

- Accelerate antibody-antigen reaction
- Provide an amplified signal for immunoassay
- Read results directly by naked eyes
- Convenience
- Test without the handling of reagents
- One-step assay



Gold nanoparticle colloidal suspensions (size of the gold colloids: 5 nm, 20 nm, 30 nm, 40 nm, 50 nm, 60 nm, and 80 nm)

# Comparison of the characteristics of labels commonly used in rapid tests

Feature	Gold	Silver	Carbon	Latex	Dye	Enzyme
Visibility	***	*	***	***	***	***
Sensitivity	***	*	**	**	**	***
Stability	***	***	**	**	**	*
Colors	*	—	*	***	***	**
Reproducibility	***	***	*	*	*	**
Scale-up	***	***	**	*	*	*
One step	***	***	***	***	***	—
Multianalyte detection	***	***	**	**	**	**
Clean result	***	***	*	**	*	*
Ease of preparation	***	***	**	***	**	**
Ease of use	***	***	**	***	**	*
Adaptability	**	**	**	**	**	***
Low cost	***	***	***	***	***	**
* Limited application						
** Acceptable for some applications						
*** Outstanding and applicable to most tests						

# Synthesis of colloidal gold

## Nanogold colloidal particle

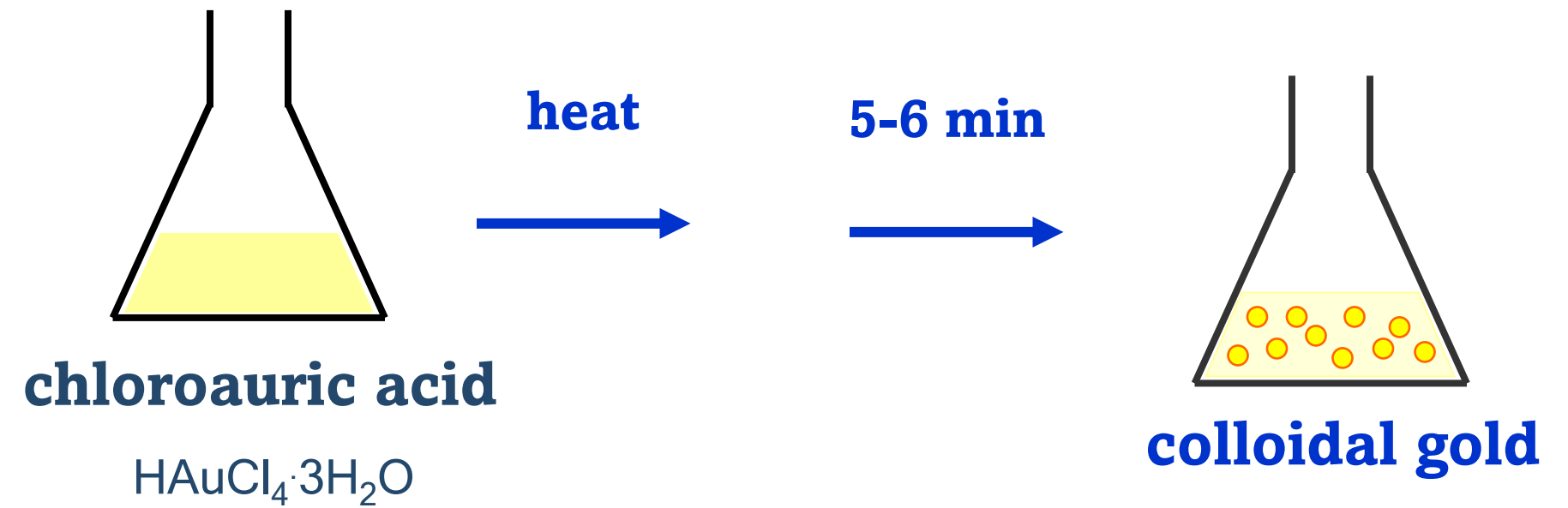
- Stable and easy to use
- Absorb at different wavelengths

Formation of antibody-colloidal gold conjugates

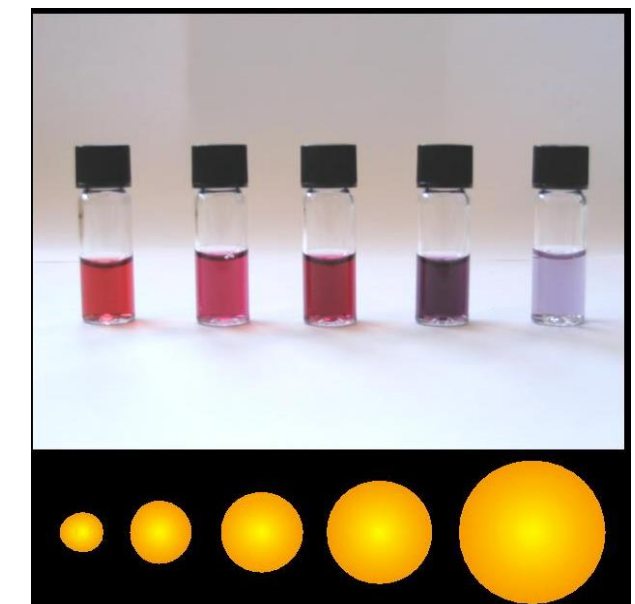
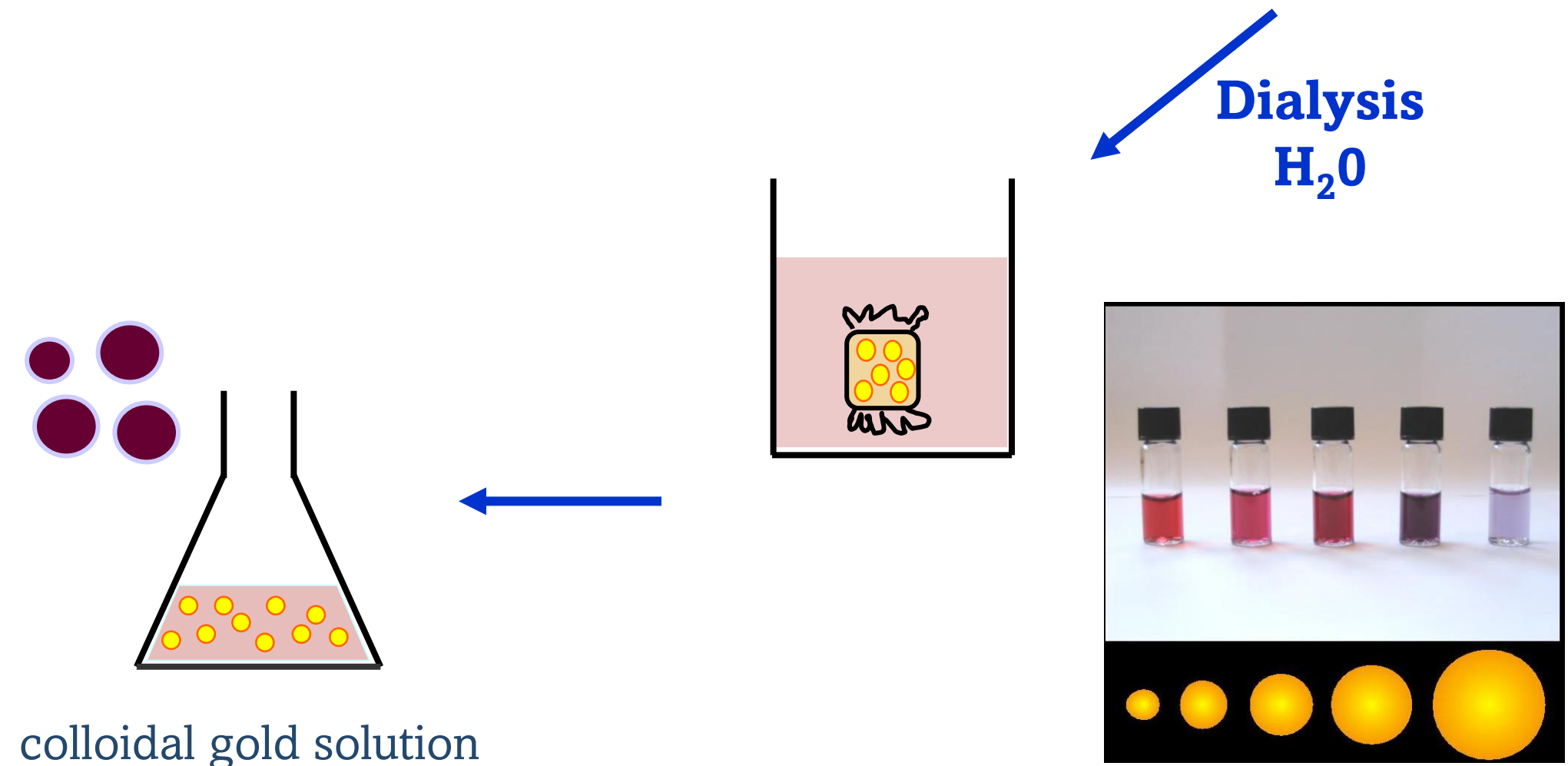
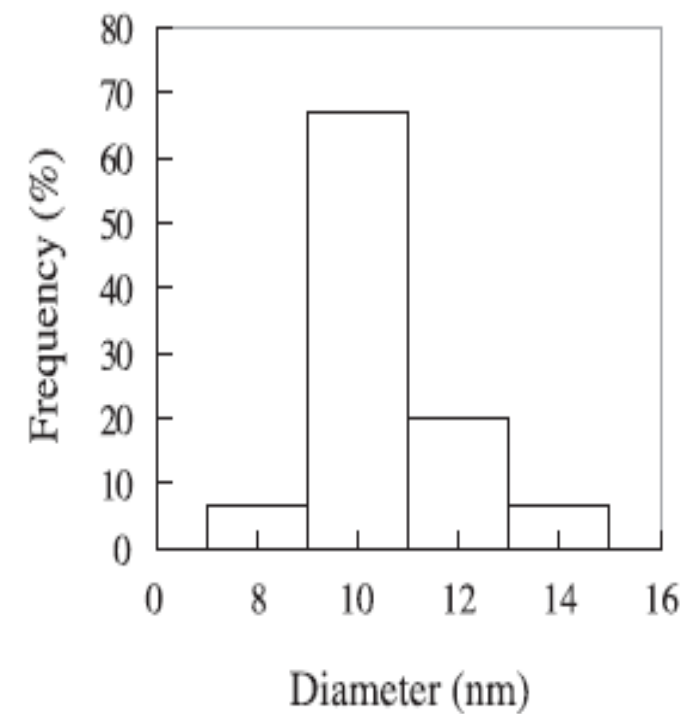
: Measured the size distributions

Transmission Electron Microscopy(TEM)

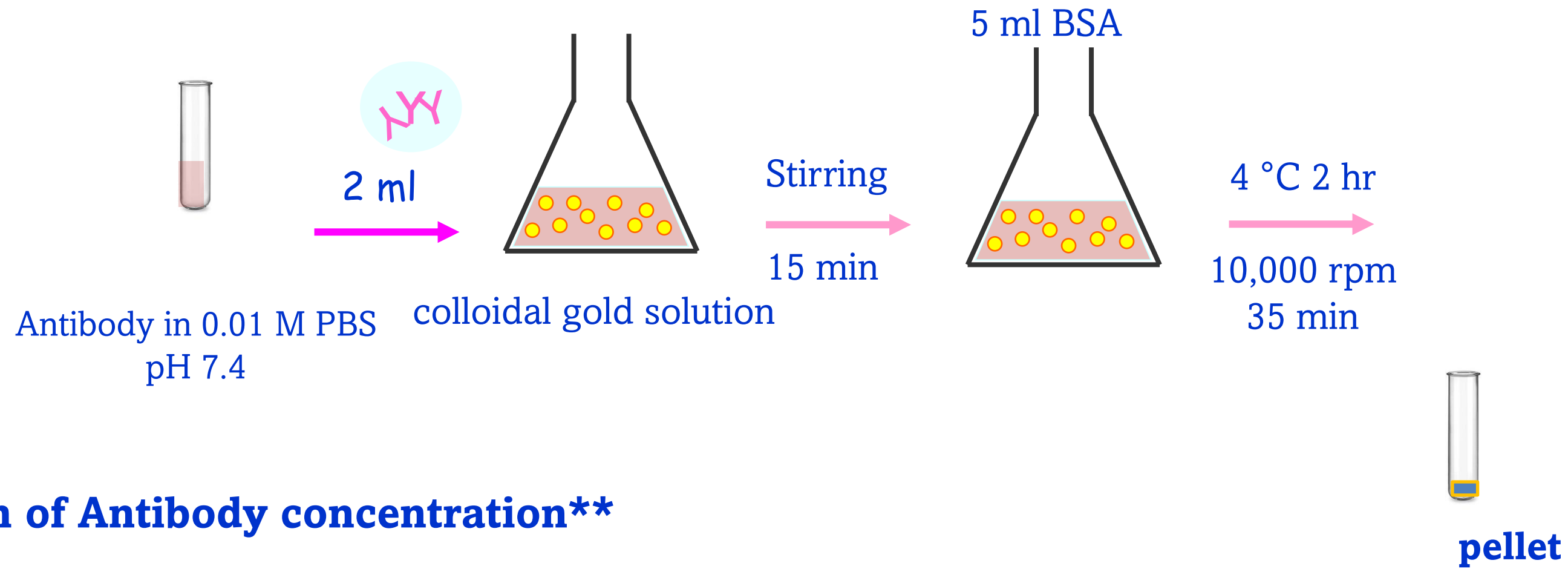
UV-vis spectroscopy



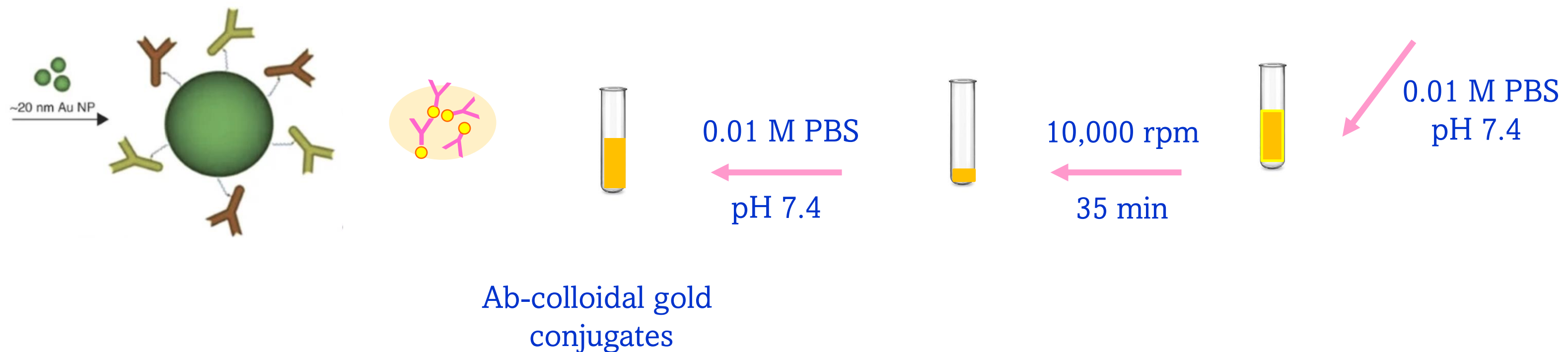
Average diameter  $10.7 \pm 1.5$  nm



# • Formation of antibody-colloidal gold conjugates



## \*\*Optimization of Antibody concentration\*\*



# Lateral flow strip test components

- Sample pad
- Conjugate pad
- Analytical pad
- Absorption
- Backing card

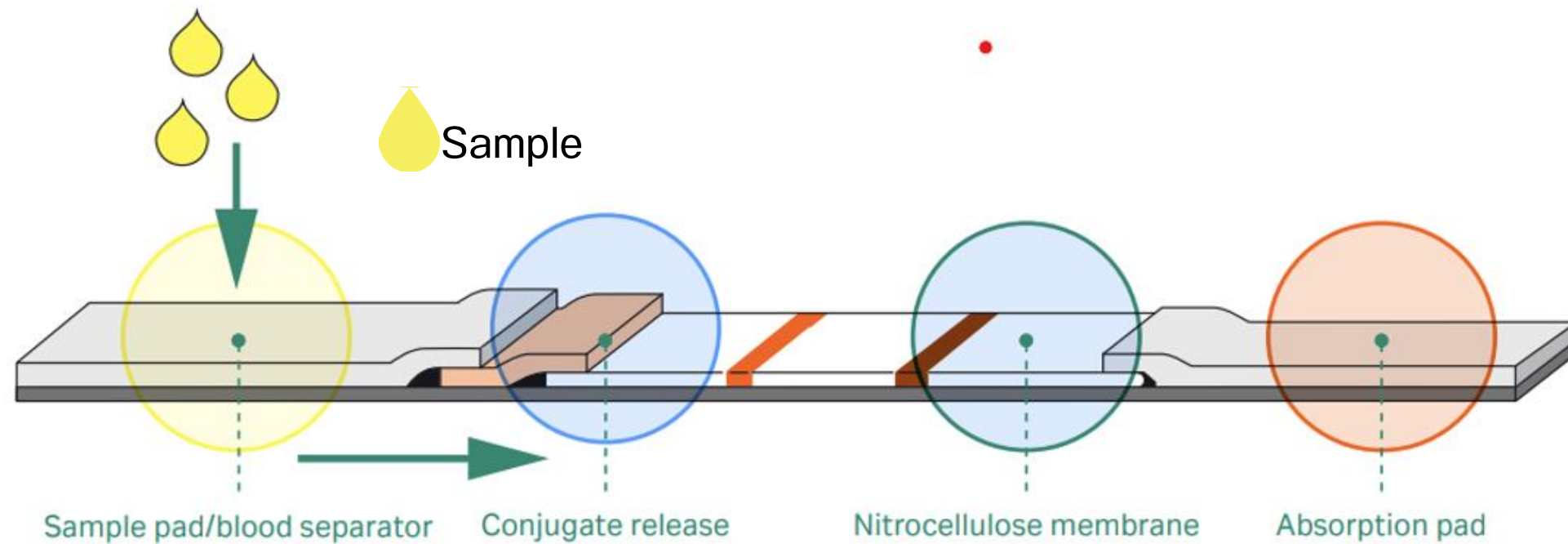


Fig. 2 Drawing of a strip test immunoassay

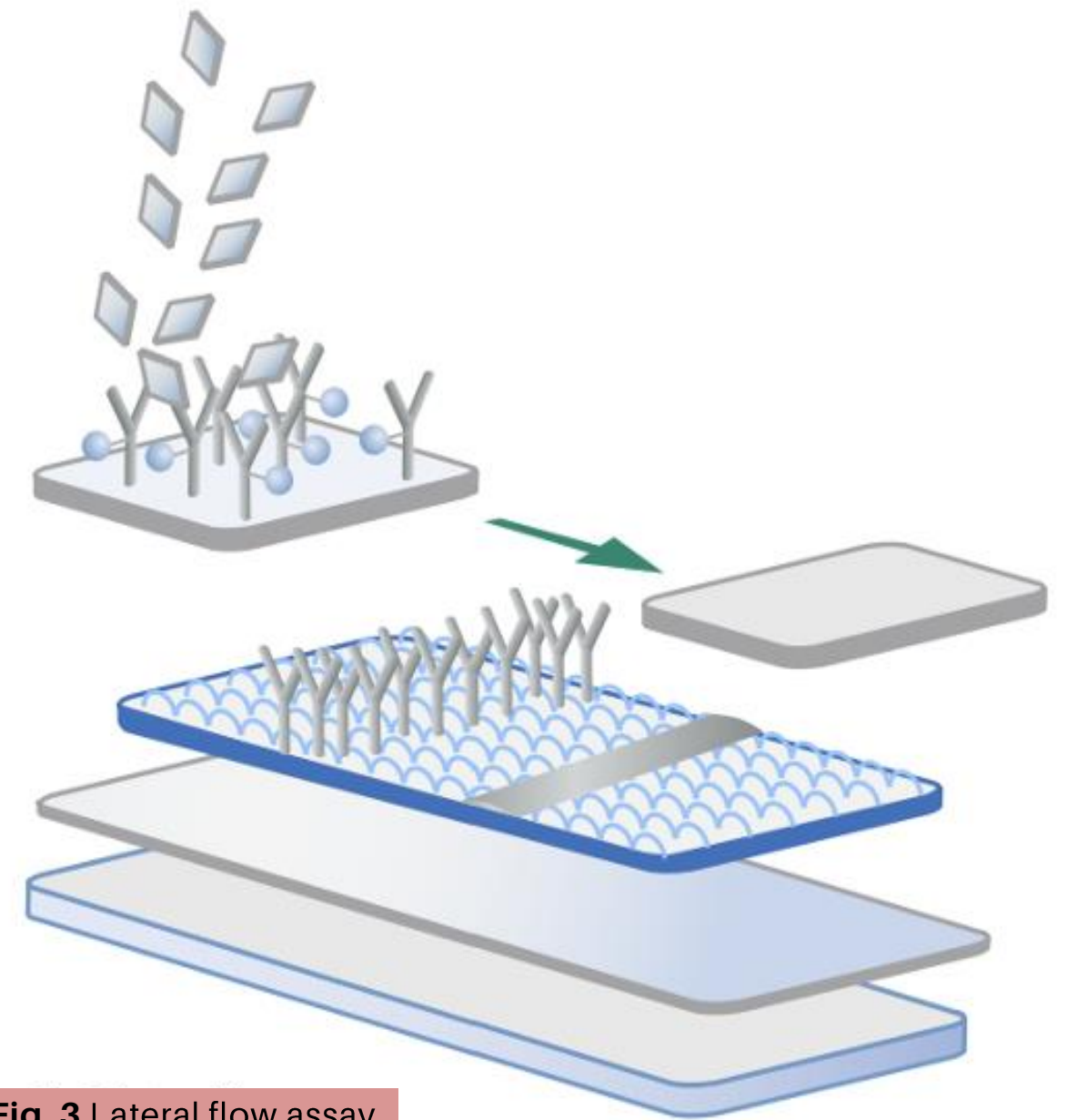


Fig. 3 Lateral flow assay

# SAMPLE PAD

Choosing a sample pad with the properties are suitable for your sample (volume and type of samples)

## Chemical properties

- pH
- Ion strength

## Physical properties

- Sample or target size
- Viscosity
- Volume

Whatman™ sample pads are available in a wide range of thicknesses, absorbencies and wicking rates.

Volume per sq cm	Grade	Properties	Thickness (µm @ 53 kPA)	Wicking rate (s/4 cm)	Water absorption (mg/cm <sup>2</sup> )
< 50 µL	<u>CF1</u>	A thin, smooth-surfaced cotton linter paper with a liner flow rate, suitable for small volumes.	176	187	16
50–100 µL	<u>CF3</u>	A medium thick cotton linter paper, originally used for separation of inorganic compounds. Larger sample volume than CF1.	322	161	31
100–150 µL	<u>CF4</u>	A medium thick cotton linter paper with acid treatment to improve wet strength and reduce trace impurity content. Similar weight and thickness to CF3 with faster wicking.	482	65	46
> 150 µL	GR470	Untreated bound cellulose suitable for whole blood or serum. Performs well with one or two drops of whole blood.	840	77	78
> 50 µL	<u>Standard 14</u>	Untreated bound glass fiber for faster flow than cotton with lower sample retention. Higher absorption capacity than Standard 17.	355	23.1	50.9
> 50 µL	<u>Standard 17</u>	Untreated bound glass fiber for faster flow than cotton with lower sample retention. Greater tensile strength than Standard 14.	370	34.5	44.9
> 50 µL	<u>GF/DVA</u>	Untreated bound glass fiber particularly suitable for saliva samples and raw milk samples.	785	28.2	93
> 50 µL	VF2	Bound glass fiber used as a single or multiple layers for separation.	785	23.8	86.2

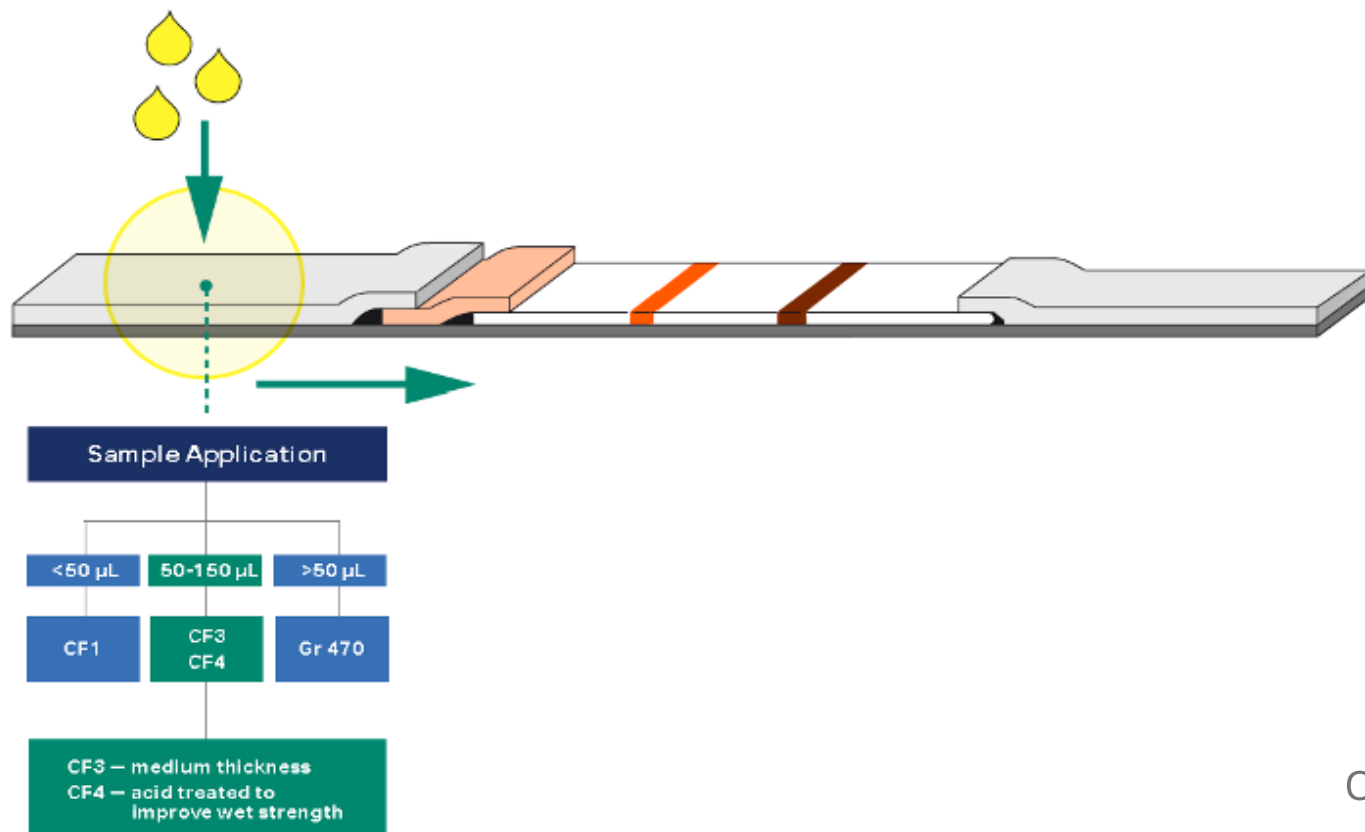
Ref: <https://cdn.cytivalifesciences.com/api/public/>

\*\* Define sample pad and pretreatment

# SAMPLE PAD

## Technical properties

Product	Material	Properties	Thickness ( $\mu\text{m}$ @ 53 kPA)	Wicking Rate (s/4 cm)	Water absorption ( $\text{mg}/\text{cm}^2$ )
CF1	100% cotton linter	Light, thin grade suitable for small volume	176	207.3	18.7
CF3		Medium weight	322	174.3	34.6
CF4			482	67.3	49.9
470			840	77	78
Standard 14	Bound glass fiber	Faster flow than cotton, with lower sample retention	355	23.1	50.9
Standard 17			370	34.5	44.9
GF/DVA		Works well with saliva samples and can act as a blood separator as well	785	28.2	93
LF1		Works well with whole blood or serum samples and can act as a blood separator as well	247	35.6	25.3
MF1			367	29.7	39.4
VF2			785	23.8	86.2



## BLOCKING AGENTS FOR SAMPLE PAD

Blocking agents can be soaped to reduce the non-specific binding of the analyte or detection reagents to the reaction membrane such as

- BSA
- Detergent >> Tween 20/Tween 80
- Buffer substance



# CONJUGATE RELEASE PAD

- Zone for immobilized Specific Mab conjugated Gold
- First place of conjugate and target sample interaction
- Glass fiber is a popular material.
- Conjugate buffer is used as a preservative (especially Carbohydrate).

\*\* Requires equipment to spray

## Conjugate Release Pad: Pretreatment

- Buffer substance
- Non-ionic surfactant
- Protein and/or polymer (reduce non-specific binding)
- Need Sugar (Trehalose or Sucrose)

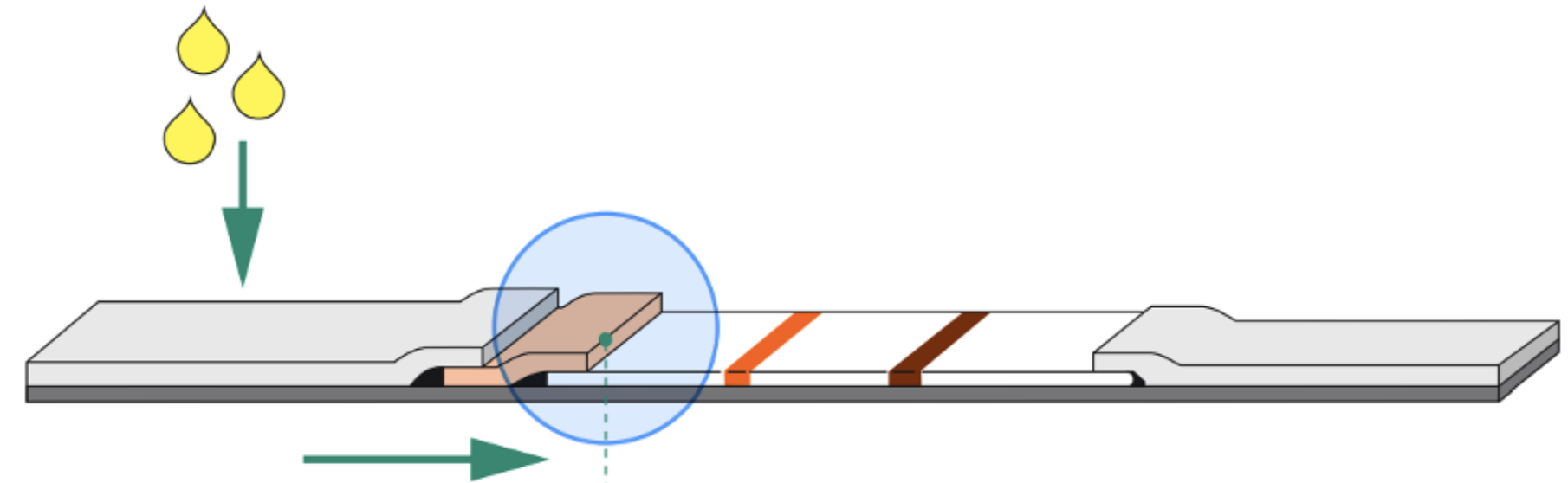


Fig 7. Conjugate release selection tree.

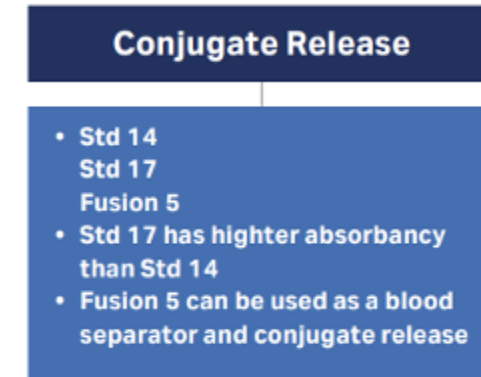


Fig. 4 Conjugate release pad

### Technical properties

Grade	Thickness (µm @ 53 kPA)	Wicking rate (s/4 cm)	Water absorption (mg/cm <sup>2</sup> )	Percent release of gold conjugate (after 90 s)
Standard 14	355	23.1	50.9	75
Standard 17	370	34.5	44.9	75
Fusion 5	370	43.9	42.3	>94

Ref: <https://www.cytivalifesciences.com/en/us/solutions/lab-filtration/knowledge-center/conjugate-release-pads>



# ANALYTICAL PAD: MEMBRANES

Analytical pad Selection to Strip test development generally is selected by

- Capillary flow times
- Surfactant contents



Fig 1. Basic set-up of membrane capillary flow time measurement. Strip width is 1 cm, strip length 4.5 cm, with triangular marks at 4 cm strip length. The time the liquid (water) needs to reach these marks is taken and documented. Water volume is 100  $\mu$ L.

## Base properties of Cytiva lateral flow membranes

Table 1. Summary of Cytiva membrane properties

Membrane grade	Backed/unbacked	Capillary flow time (seconds/4 cm)	Relative surfactant concentration	Use recommended for sample	Additional information
FF80HP	Backed	60–95	Low	Serum, plasma	In general, used for high viscosity samples, should be blocked in a separate manufacturing step
FF120HP	Backed	90–140	Low	Diluted serum, plasma, urine	Membrane of choice for lateral flow arrays. Requires blocking in a separate manufacturing step
FF170HP	Backed	140–200	Low	Low viscosity samples, urine, water	Is also being used for high viscosity samples and capture reagents with slow on-rates
FF80HP Plus	Backed	60–95	Medium	Serum, plasma, milk	
FF120HP Plus	Backed	90–140	Medium	Diluted serum, plasma, urine	
FF170HP Plus	Backed	140–200	Medium	Low viscosity sample, urine, water	
Immunopore RP	Backed	90–150	High	Serum, plasma, urine	
Immunopore FP	Backed	140–200	High	Diluted serum, urine water	
Immunopore SP	Backed	200–300	High	Low viscosity samples in general	Membrane of choice for capture reagents with very slow rates. It will result in longer test duration times
Prima 40	Backed	38–52*	Very high	Very high viscosity samples, milk	Originally developed for test systems aiming for the detection of whole cells in lateral flow tests
AE 100	Unbacked	90–120	Medium	Serum, plasma	
AE 99	Unbacked	120–160	High	Urine, diluted serum	
AE 98	Unbacked	140–200	Medium	Urine, low viscosity samples	

\*The capillary flow time in Prima 40 is determined by a method that is different from the one described in Figure 1

Please note that the use recommendations only refers to first experiments. It may be that a developer can or must use a membrane with a faster or slower capillary flow time, depending on the kinetic properties of the reagents that are to be used in the test system. Whole blood has not been listed in the table as a sample liquid since the blood cells in lateral flow tests are usually retained in the sample pad, and the liquid that runs through the membrane will be serum or plasma even in that case.

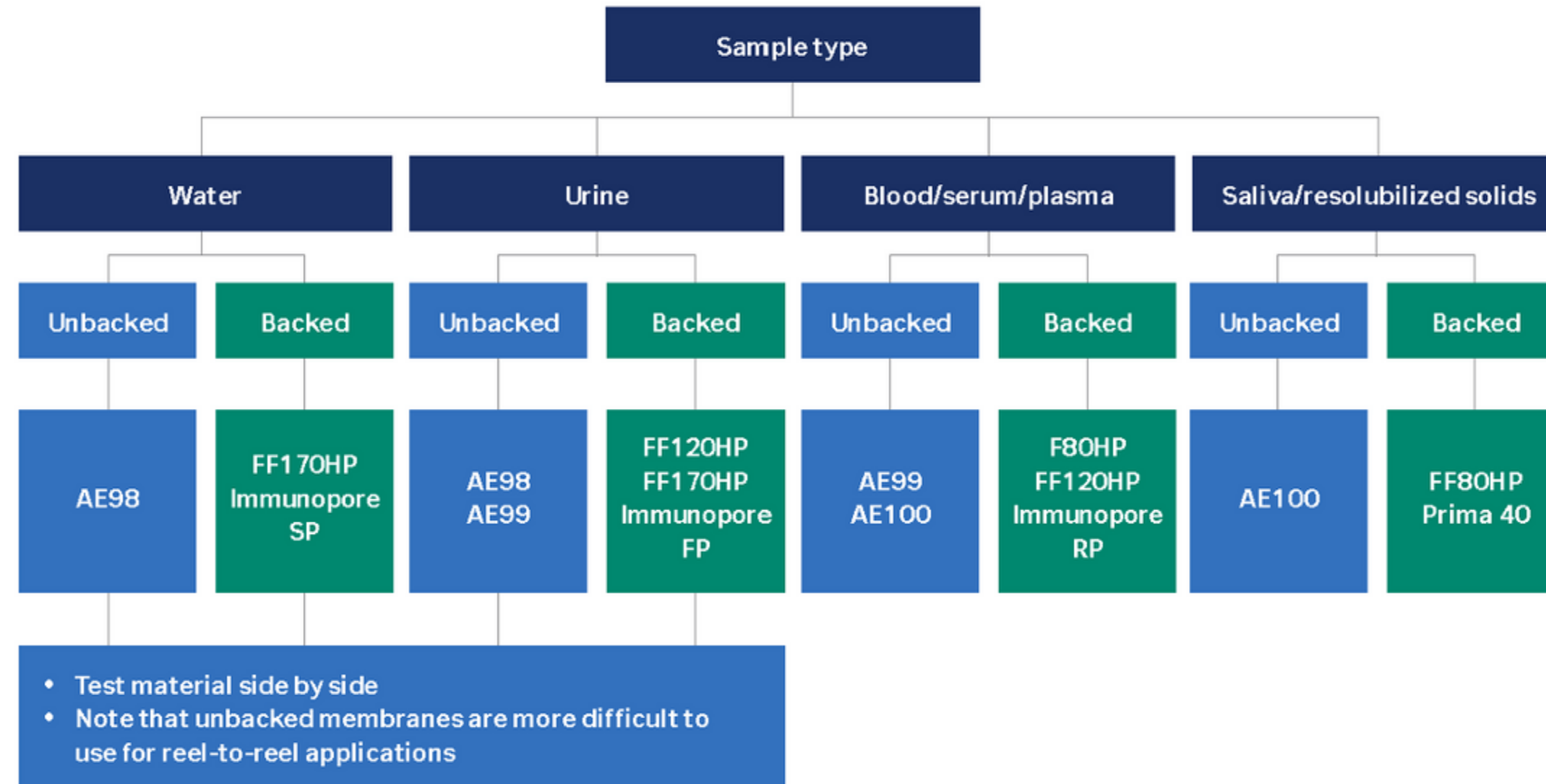
## Capillary flow times

Ref: <https://www.cytivalifesciences.com/en/us/solutions/lab-filtration/knowledge-center/conjugate-release-pads>

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# ANALYTICAL PAD: MEMBRANES

## Membrane selector according to sample type

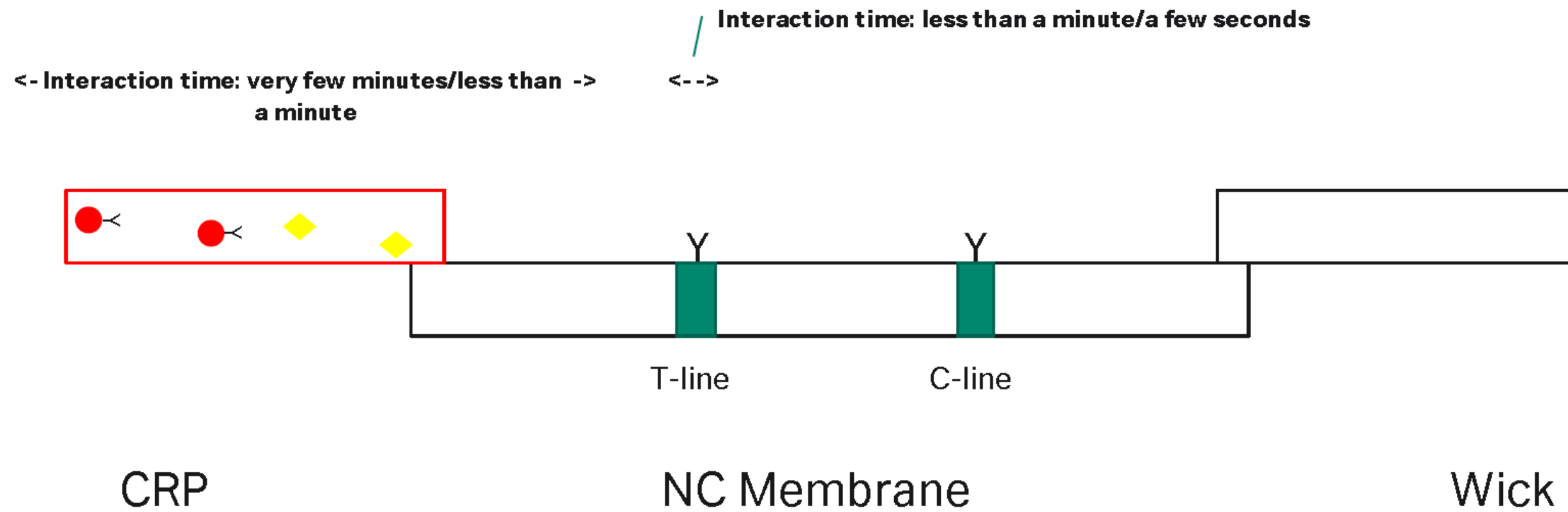


**Table 2:** Comparison between backed and unbacked membrane features

Backed membrane	Unbacked membrane
<ul style="list-style-type: none"> <li>• Increased mechanical strength of the membranes, simplifying use in reel-to-reel machines</li> <li>• Direct contact is prevented between the nitrocellulose material and the adhesive from the lamination card where the test elements are mounted</li> </ul>	<ul style="list-style-type: none"> <li>• Enables assay suitability tests of both air and belt side of the membrane</li> </ul>

# Interaction times

- ◆ Target molecule
- Detector conjugate



**ISSUES WITH SLOW MEMBRANES:**  
False positive signals, high background

**ISSUES WITH FAST MEMBRANES:**  
False negative results

# ABSORBENT PAD

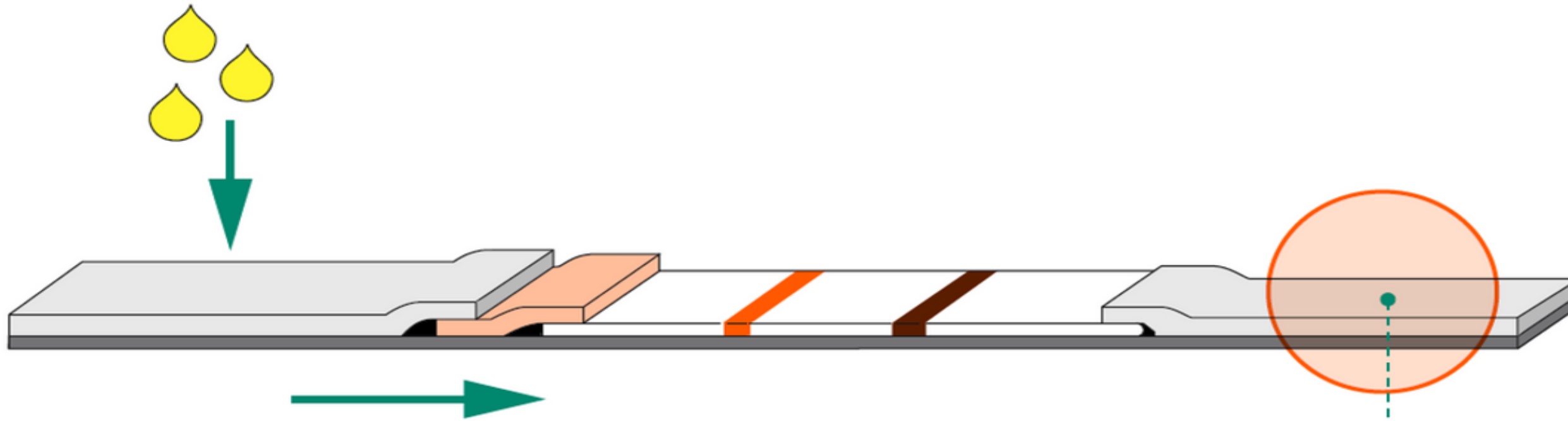
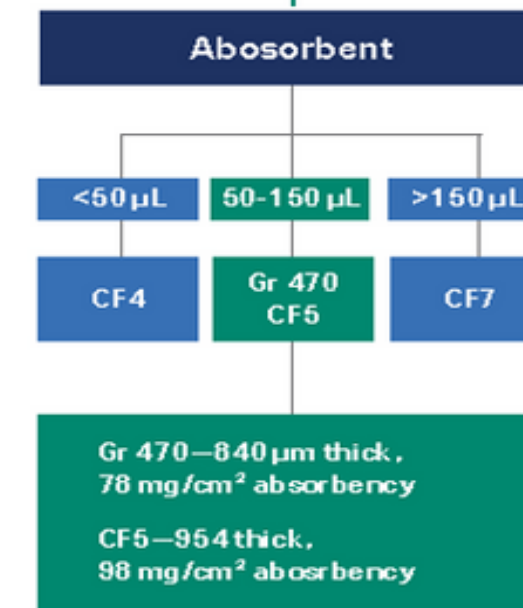
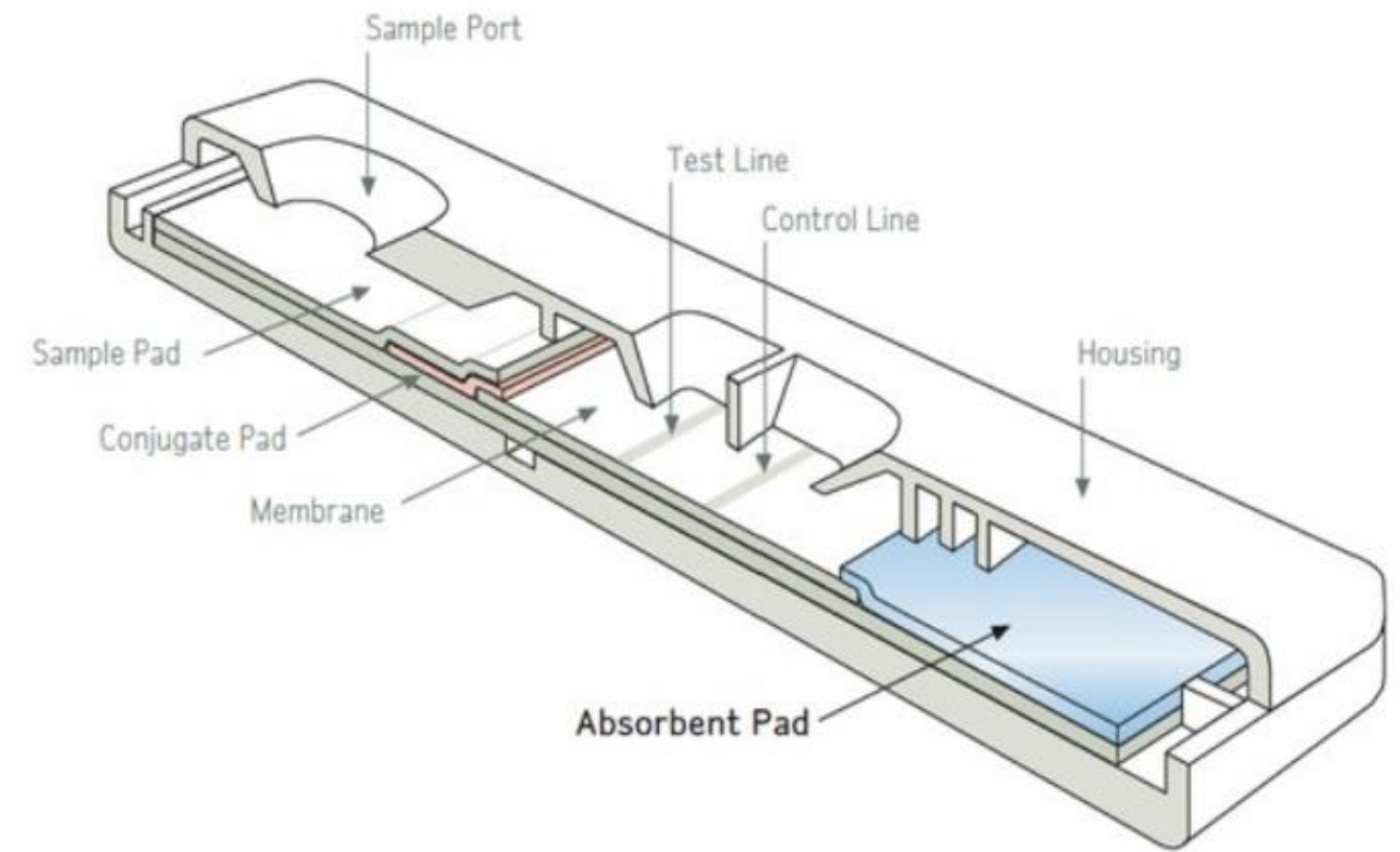


Fig 12. Absorption pads selection tree.



# ABSORBENT PAD

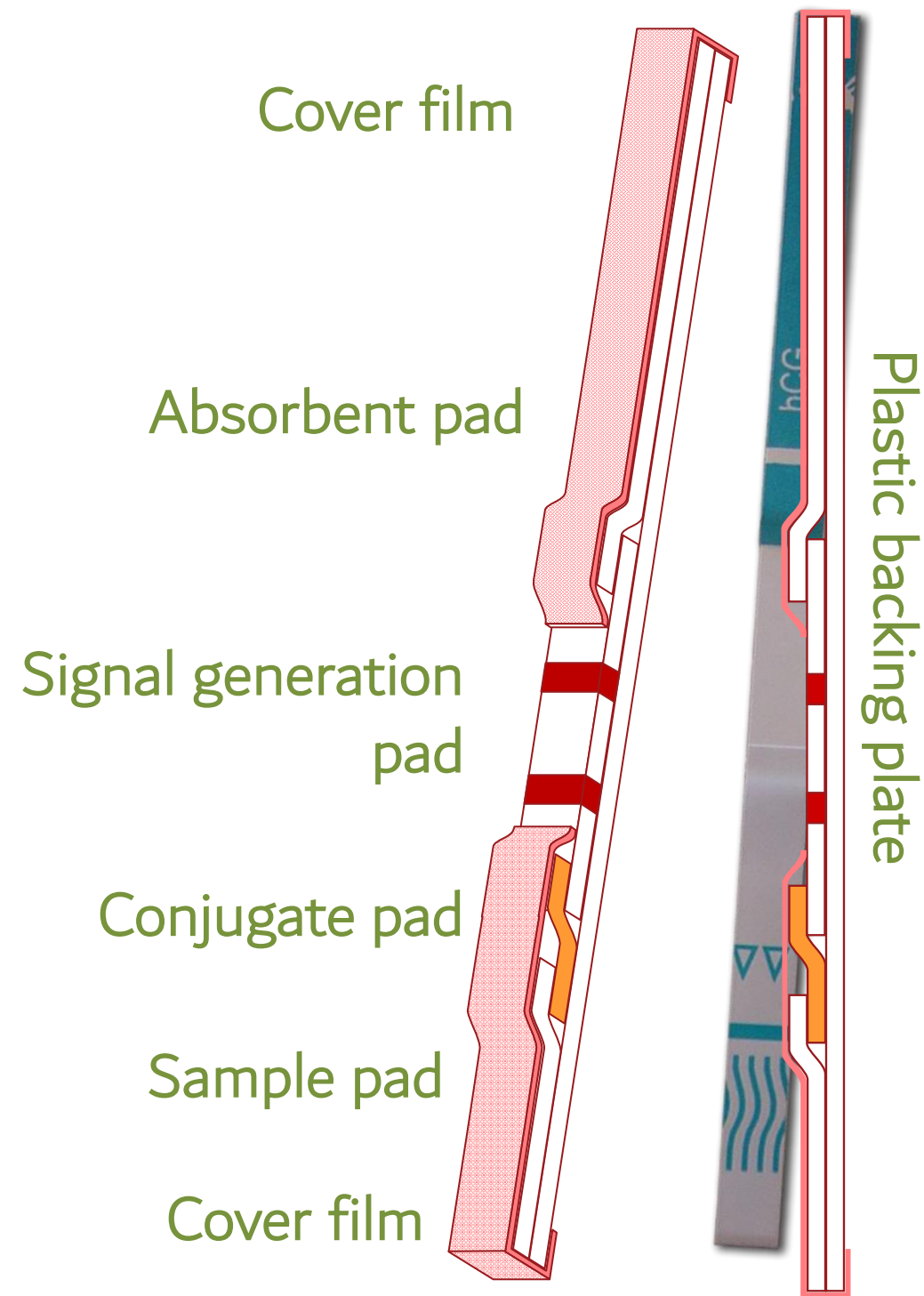
- To wick the fluid through the membrane
- To collect the processed liquid



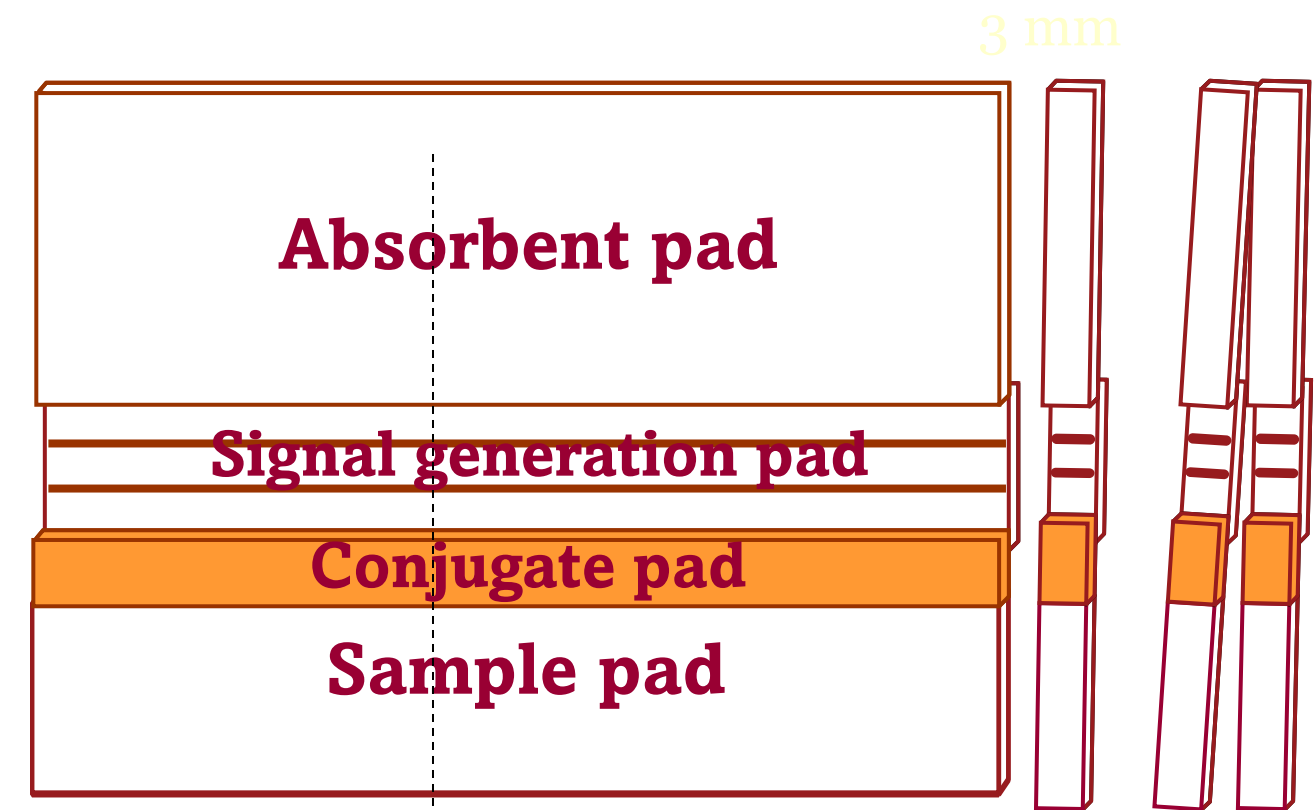
Ref. Clayton, Katherine Noel. "Comparing Anti-VEGF Antibodies and Aptamers on Paper Microfluidic-based Platforms." PhD diss., California Polytechnic State University, 2012.

Grade	Properties	Thickness (um @ 53kPA)	Wicking rate (s/4cm)	Water absorption (mg/cm <sup>2</sup> )
<u>CF3</u>	A medium thick cotton linter paper, originally used for separation of inorganic compounds. Larger sample volume than CF1.	322	174.3	34.6
<u>CF4</u>	A medium thick cotton linter paper with acid treatment to improve wet strength and reduce trace impurity content. Similar weight and thickness to CF3 with faster wicking.	482	67.3	49.9
<u>CF5</u>	100% cotton linter can handle medium/high volumes	954	63.3	99.2
<u>CF7</u>	100% cotton linter can handle medium/high volumes	1873	35	252.3
GR470	Untreated bound glass fiber suitable for whole blood or serum.	840	77	78

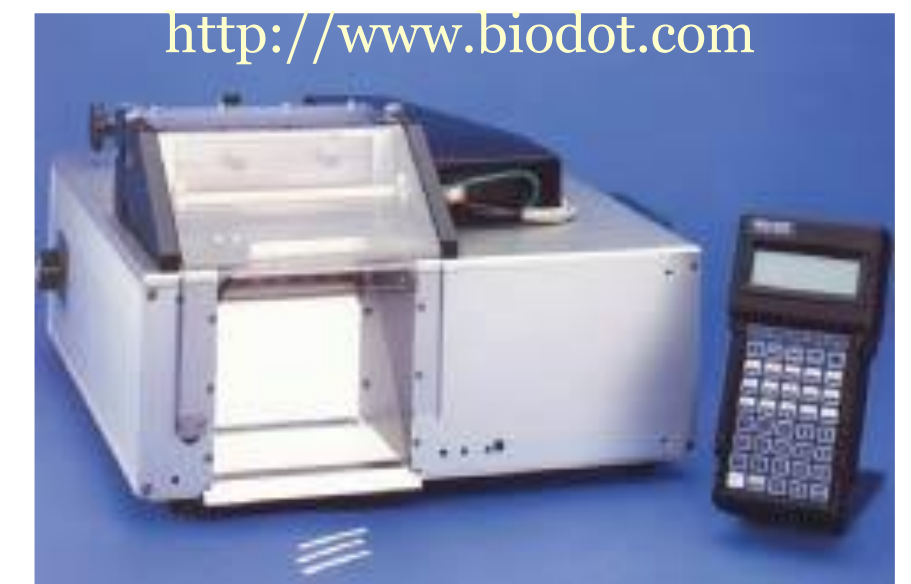
# METHODS



# ASSEMBLING

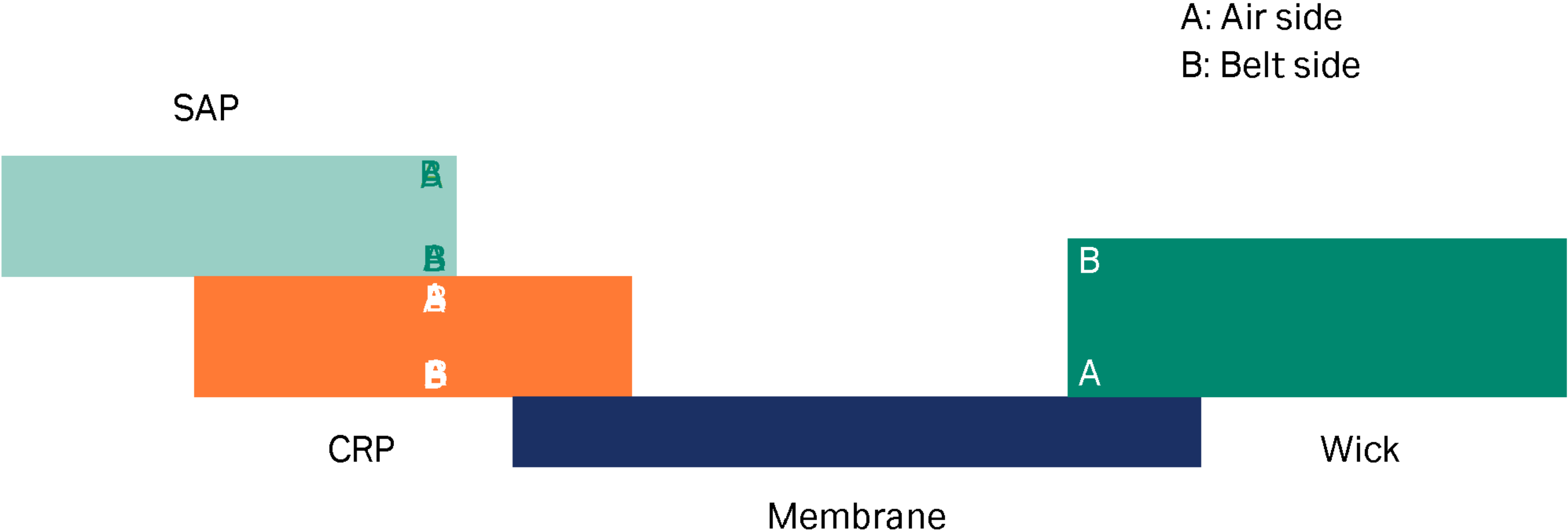


- **Cut strip using a CM4000 cutter**
- **Store at 4°C**



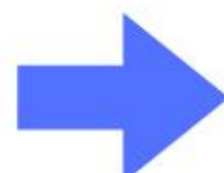
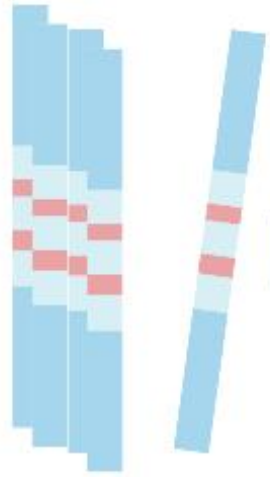
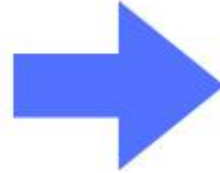
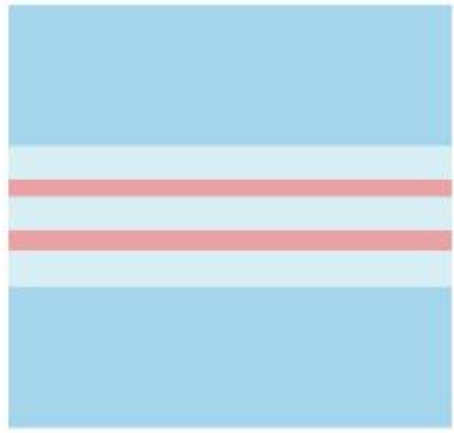
# Warning !!

## Paper orientation in lateral flow tests





# ASSEMBLING



Assemble >> all components of the strip test

Cut to 4 cm

Put to the cassette

Put into aluminum foil

Pack to a box



# Information/Instruction for use (IFU)

## Information needs

### Manufacturer information

Produce name :

Application:

Country of Origin:

Code of product:

Manufacturer address:

### Product Description

1. Intended use
2. Summary and explanation
3. Device Description and Features
4. Medical device contents
5. How to storage
6. Shelf-life
7. Test procedure
8. Internal Quality Control
9. Indications
10. Warnings and Precautions (this information should be related with risk analysis report)
11. Limitations
12. Materials and components
13. Other relevant specifications

01/08/2024

### Performance characteristics

1. Test Verification
2. Cut off value
3. Cross-reactivity
4. Interference substance
5. Precision
6. Hook effect

# Immunochromatographic strip test Production

